

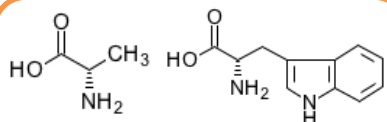
Bases de données en métabolomique

Christophe Junot

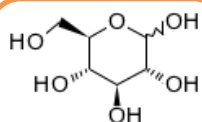
**CEA/Laboratoire d'Etude du Métabolisme des Médicaments
CEA-Saclay (iBiTec-S)
christophe.junot@cea.fr**

Metabolites and metabolome

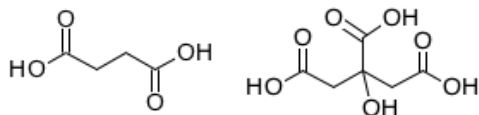
Primary metabolites



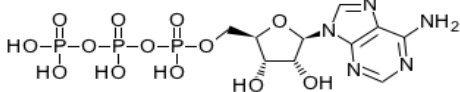
Aminoacids



Sugars



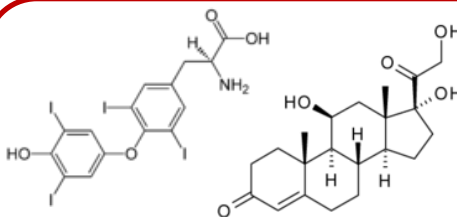
Organic acids



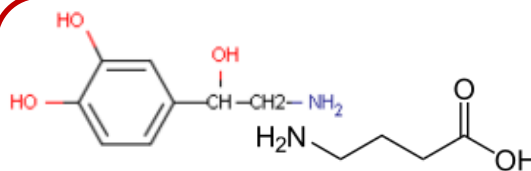
Nucléotides

(...)

Secondary metabolites



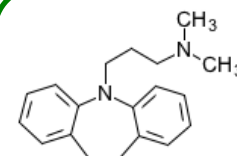
Hormones



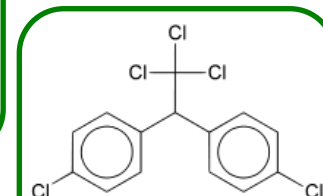
Neurotransmitters

(...)

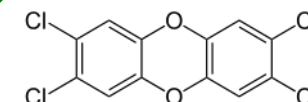
Xenobiotics



Drugs



Pesticides



Pollutants

(...)

Food / drinking

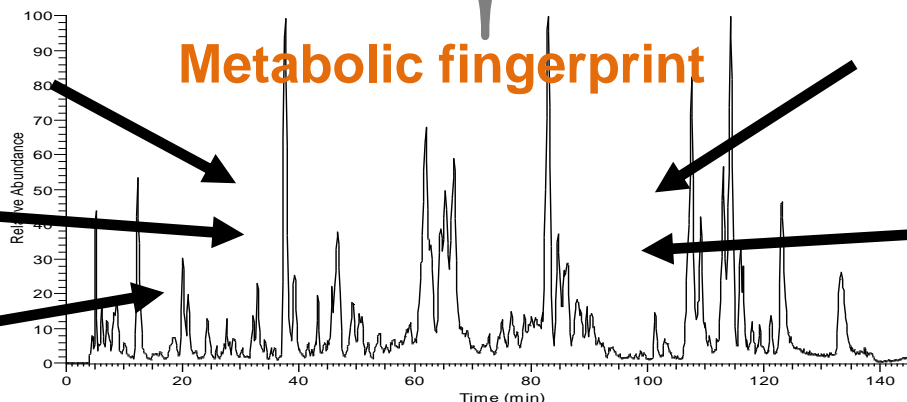
Metabolic fingerprint

Central metabolism

Gut microbiota

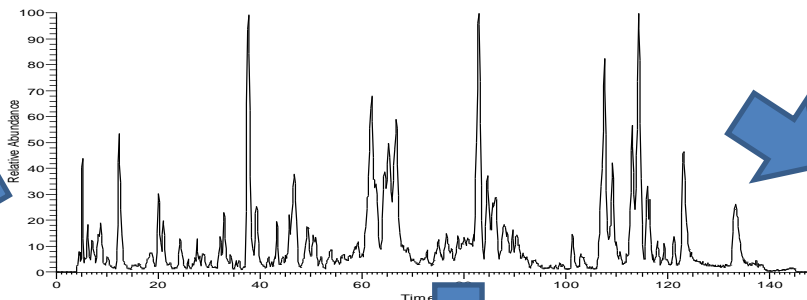
**Environment :
xenobiotics
(pollutants, drugs...)**

Pathology



How to detect metabolites in biological media?

Metabolic fingerprint



NMR

- Simple, non invasive
- Rapid
- Robust: analysis of large series of samples
- But:
- Limited sensitivity

GC-EI-MS

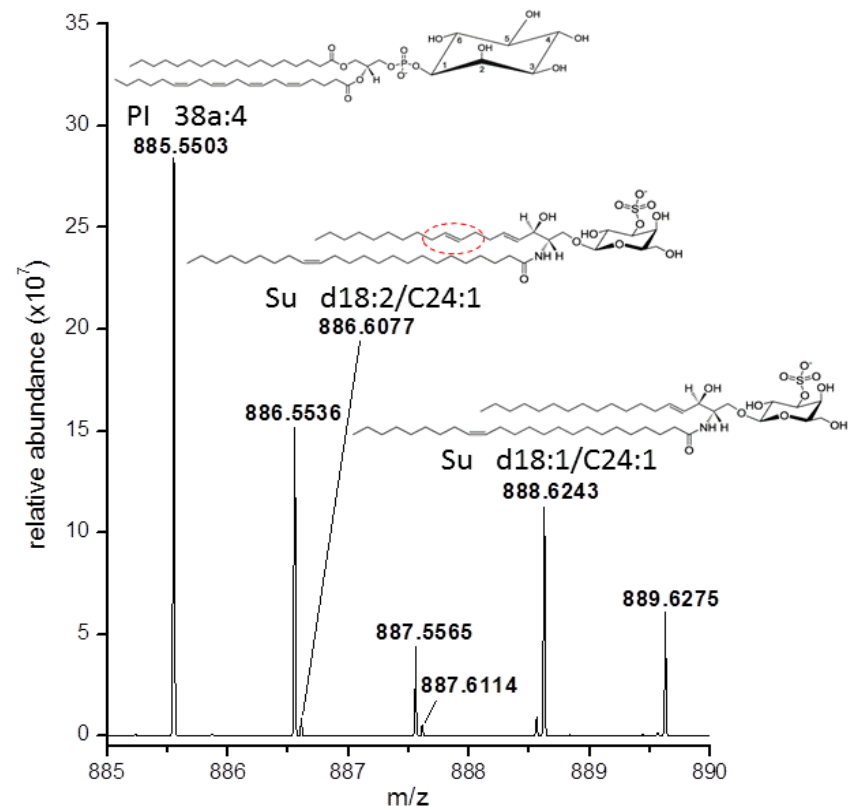
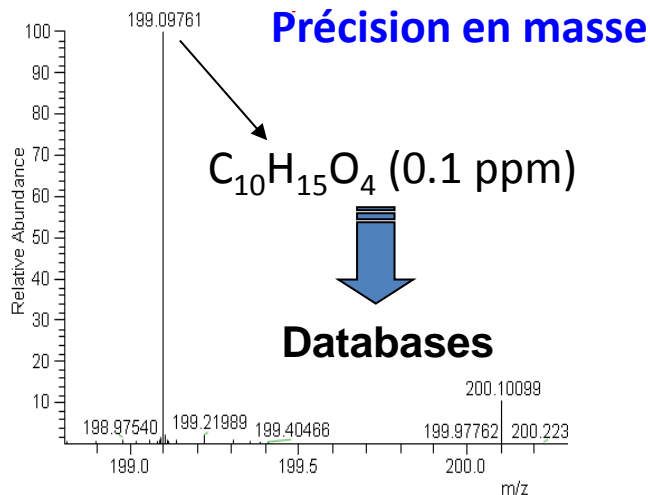
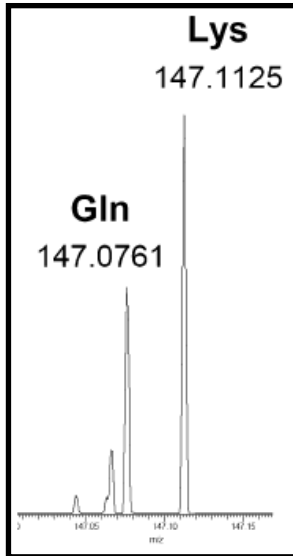
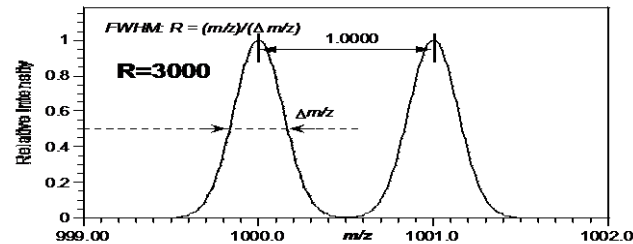
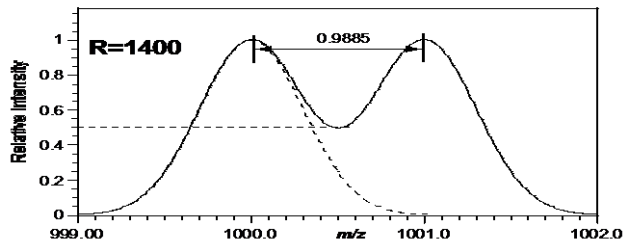
- Sensitive
- Reproducible
- Spectral libraries
- But:
- Chemical derivatization of non volatile compounds
- Issue of thermolabile compounds

LC-API-MS

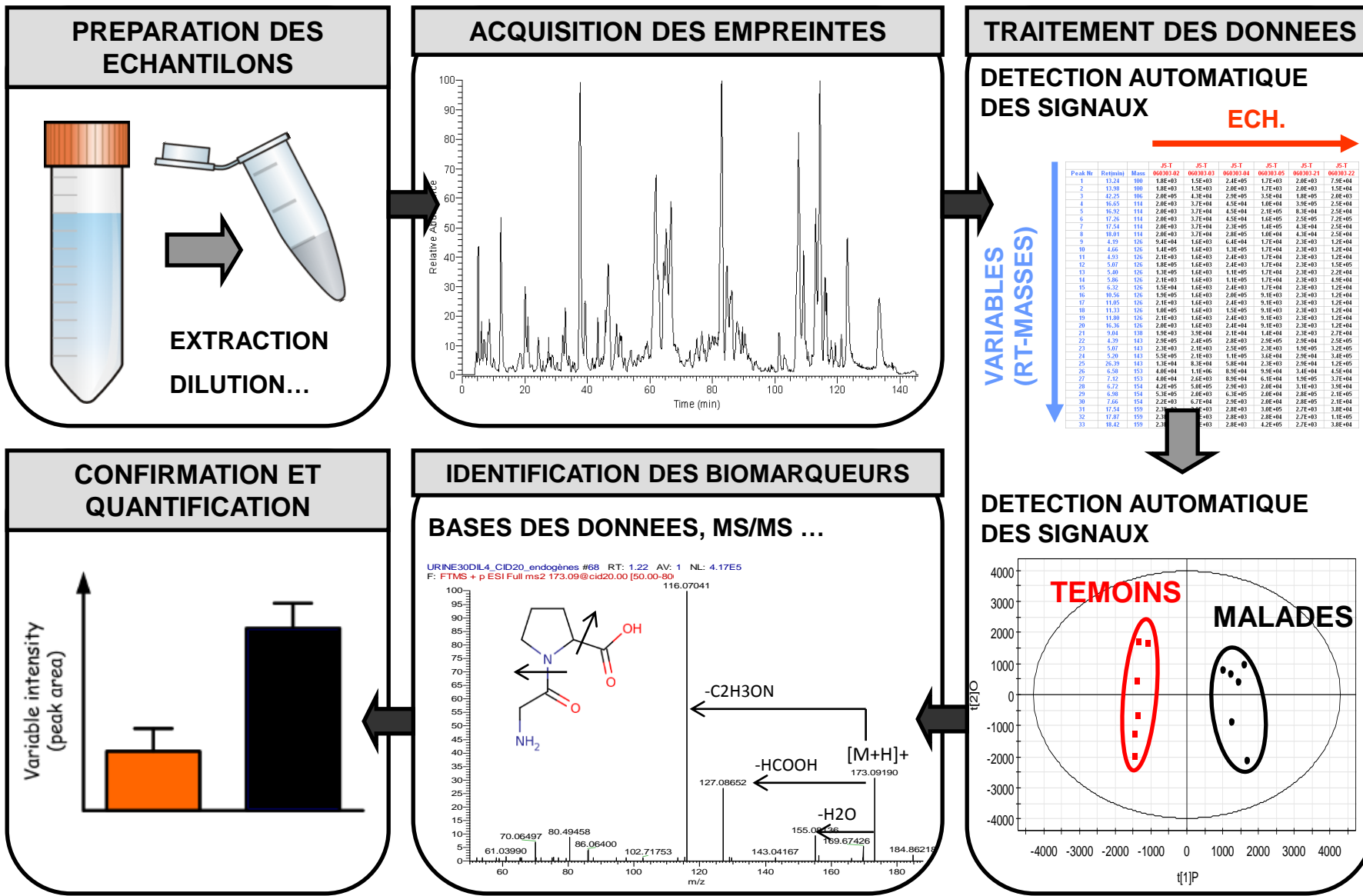
- Molecular mass of intact compounds
- Analysis of thermolabile compounds
- sensitive
- But:
- Poor inter-platform reproducibility

MS à haute résolution: détecter plus de métabolites et les identifier plus facilement

Résolution en masse



Déroulement d'une analyse métabolomique

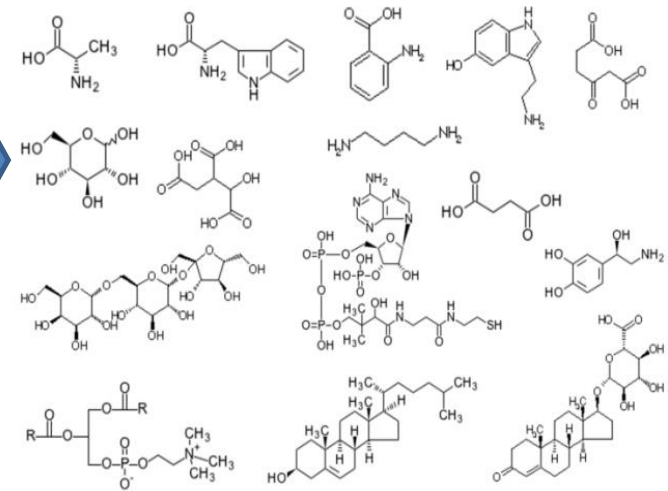
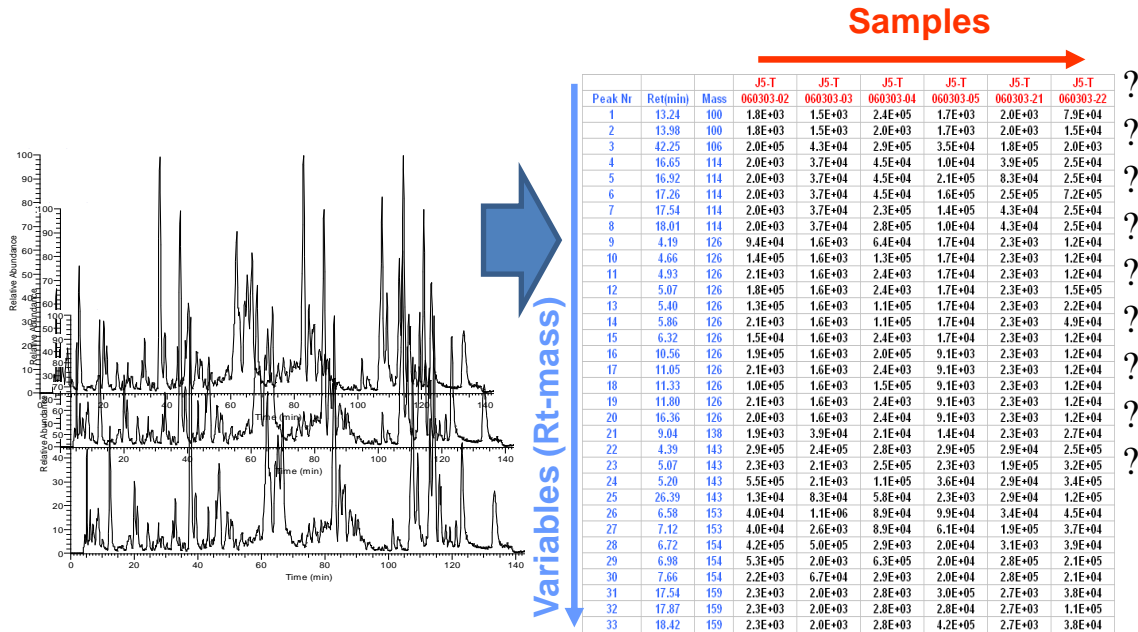


Spectral databases

database		thematic	Conception / URL	Instrument
NIST	⊙	general	National institute for standard and technology (USA) www.nist.gov/srd/nist1a.htm	GC/MS
Fiehn Library	●	general	Fiehn Laboratory Univ California Davis – Genome center http://fiehnlab.ucdavis.edu/Metabolite-Library-2007	GC/MS
Golm	●	plant	Max Planck Institute for Molecular Plant Physiology (Germany) csbdb.mpimp-golm.mpg.de	GC/MS
HMDB	●	human metabolites	Department of Computing Science, University of Alberta (Canada) www.hmdb.ca/extrIndex.htm	NMR, API/MS/MS
Lipidmaps	●	lipidomics	LIPID MAPS Bioinformatics Core (USA) www.lipidmaps.org/data/index.html	API-MS/MS
Massbank	●	general	Keio university, university of Tokyo, Kyoto university, RIKEN plant Science center (Japan) and others www.massbank.jp	API-MS/MS
Metlin	●	human metabolites	Scripps Center for Mass Spectrometry metlin.scripps.edu	API-MS/MS
Brucker	○	general		NMRS
Madison Metabolomic Consortium database	●	general	http://mmcd.nmrfa.wisc.edu/	NMRS

● free access, ⊙ partially free access, ○ licenced

Annotation of peak lists is required to help for metabolite identification



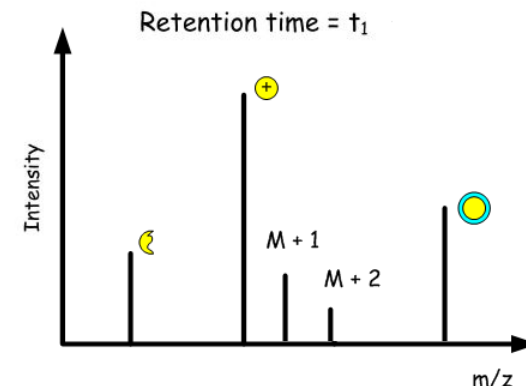
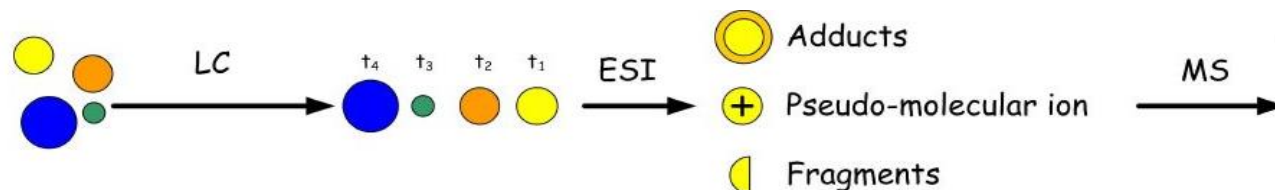
Few thousands of variables...

...Few hundreds of metabolites ??

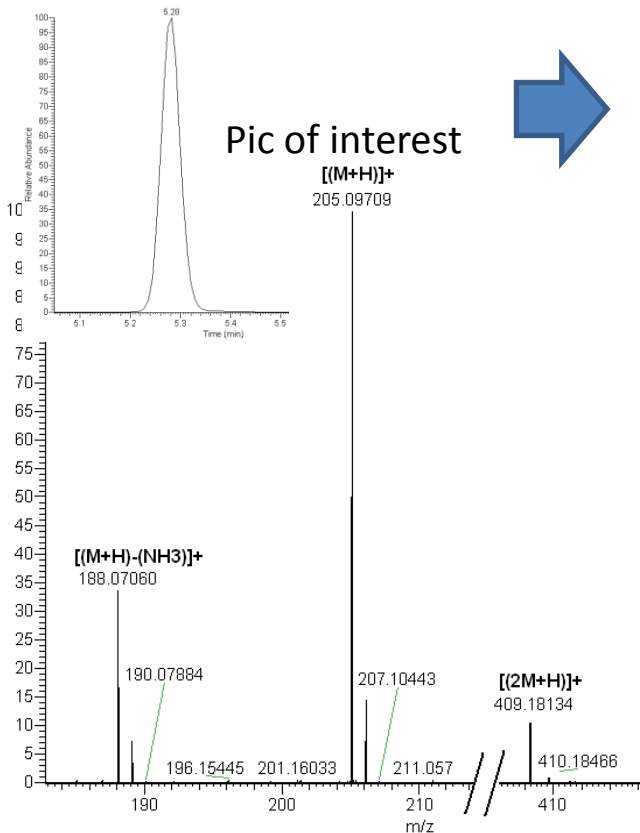
- Chemical and biochemical databases: **KEGG** (www.genome.jp/kegg), **Metlin** (www.metlin.scripps.edu), **HMDB** (www.hmdb.ca)
- spectral databases

The relevance of a spectral database

One molecule = several ions



Pic of interest



Automated detection of ions, list of annotated features

M/Z	RT	Formula	Compound	Attribution	Annotations (HMDB, KEGG, METLIN)
188.0709	5.28	C ₁₁ H ₁₀ NO ₂	Tryptophan	[(M+H)-(NH ₃)] ⁺	Deethylatrazine 3-amino-2-naphthoic acid Indoleacrylic acid
189.0757	5.28	C ₁₀ [¹³ C]H ₁₀ NO ₂	Tryptophan	[(M+H)-(NH ₃)] ⁺ (13C)	Ethyl Oxalacetate
190.0787	5.28	C ₉ [¹³ C] ₂ H ₁₀ NO ₂	Tryptophan	[(M+H)-(NH ₃)] ⁺ (13C ₂)	
205.0975	5.28	C ₁₁ H ₁₃ N ₂ O ₂	Tryptophan	[(M+H)] ⁺	Tryptophan ethotoin Vasicinol Idazoxan Nirvanol
206.1010	5.28	C ₁₀ [¹³ C]H ₁₃ N ₂ O ₂	Tryptophan	[(M+H)] ⁺ (13C)	N-Acetyl-D-fucosamine N-Acetyl-D-quinovosamine
207.1051	5.28	C ₉ [¹³ C] ₂ H ₁₃ N ₂ O ₂	Tryptophan	[(M+H)] ⁺ (13C ₂)	
409.1902	5.28	C ₂₂ H ₂₅ N ₄ O ₄	Tryptophan	[(2M+H)] ⁺	Gly Trp Phe (and isomers) Lys Met Met (and isomers)
410.1938	5.28	C ₂₁ [¹³ C]H ₂₅ N ₄ O ₄	Tryptophan	[(2M+H)] ⁺ (13C)	Tyr Leu Asp (and isomers) Ile Tyr Asp (and isomers) Val Tyr Glu (and isomers)

Development of an ESI-mass spectral database for metabolomics : methodology

Chemical library
(reference compounds)

ESI mass spectral
database

Samples

Variables
(Tr-masse)

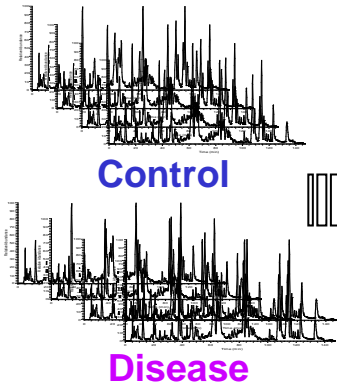
Peak #	Retention	Mass	JS-T	JS-T	JS-T	JS-T	JS-T	JS-T
1	11.23	306	1.8E+03	1.5E+03	1.7E+03	2.0E+03	2.0E+03	1.9E+04
2	12.25	306	2.0E+03	4.2E+04	2.2E+05	3.2E+04	1.8E+05	2.0E+03
3	16.63	314	2.0E+03	3.2E+04	4.2E+04	1.8E+04	3.2E+05	2.0E+04
4	16.20	314	2.0E+03	3.2E+04	4.2E+04	2.1E+05	9.2E+04	2.0E+04
5	17.24	314	2.0E+03	3.2E+04	4.2E+04	1.4E+05	2.1E+05	2.0E+05
6	16.93	314	2.0E+03	3.2E+04	2.8E+05	1.8E+04	4.2E+04	2.0E+04
7	1.17	326	9.4E+04	1.6E+03	6.4E+04	1.7E+04	2.2E+03	1.2E+04
8	1.66	326	1.4E+05	1.6E+03	1.3E+05	1.2E+04	2.2E+03	1.2E+04
9	2.17	326	1.4E+05	1.6E+03	2.4E+05	1.7E+04	2.2E+03	1.2E+04
10	2.48	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	2.2E+04
11	2.89	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	4.0E+04
12	3.30	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
13	3.71	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
14	4.12	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
15	4.53	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
16	4.94	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
17	5.35	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
18	5.76	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
19	6.17	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
20	6.58	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
21	6.99	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
22	7.40	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
23	7.81	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
24	8.22	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
25	8.63	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
26	9.04	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
27	9.45	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
28	9.86	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
29	10.27	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
30	10.68	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
31	11.09	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
32	11.50	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
33	11.91	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
34	12.32	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
35	12.73	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
36	13.14	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
37	13.55	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
38	13.96	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
39	14.37	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
40	14.78	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
41	15.19	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
42	15.60	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
43	16.01	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
44	16.42	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
45	16.83	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
46	17.24	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
47	17.65	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
48	18.06	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
49	18.47	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
50	18.88	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
51	19.29	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
52	19.70	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
53	20.11	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
54	20.52	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
55	20.93	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
56	21.34	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
57	21.75	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
58	22.16	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
59	22.57	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
60	22.98	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
61	23.39	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
62	23.80	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
63	24.21	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
64	24.62	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
65	25.03	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
66	25.44	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
67	25.85	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
68	26.26	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
69	26.67	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
70	27.08	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
71	27.49	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
72	27.90	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
73	28.31	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
74	28.72	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
75	29.13	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
76	29.54	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
77	29.95	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
78	30.36	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
79	30.77	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
80	31.18	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
81	31.59	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
82	32.00	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
83	32.41	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04
84	32.82	326	1.3E+05	1.6E+03	1.1E+05	1.7E+04	2.2E+03	1.2E+04

- Gather together signals from the same metabolite
- MSⁿ confirmation

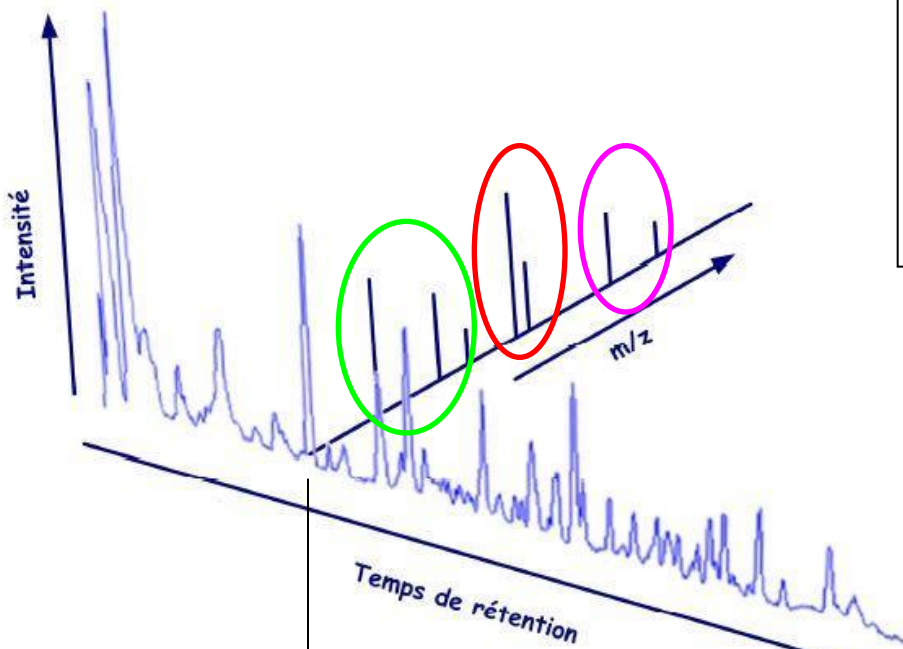
Structure dataset with multivariable analysis

Metabolomics relational database:
- Kind of biofluid
- Physiological Factors
- Disease, toxicology

Annotated peaklist



Analysis of reference compounds



One molecule = many ions :
- Pseudo-molecular and isotopes
- Adducts
- Fragments

Annot.
FIA*

M/Z	Intensity	Relative	M/Z (theo)	Delta (ppm)	RDBE	Composition	Attribution	FIA	UPLC (C8)	UPLC (C18)	HSF5
72.04431	7.7E+04	0.82	72.04439	-1.11	1.5	C3 H6 O N	[[M+H]-[GH10O3]-[H2O]]+	0	1.80	4.77	11.17
90.05482	3.6E+05	3.78	90.05496	-1.55	0.5	C3 H8 O2 N	[[M+H]-[GH10O3]]+	0	1.80	4.77	11.17
116.034	2.7E+04	0.29	116.03422	-0.22	2.5	C7 H10 O N	[[M+H]-[C2H6O3]-[H2O]]+	0	1.80	4.77	11.17
142.08585	1.0E+03	0.01	142.08626	-2.85	2.5	C7 H12 O2 N	[[M+H]-[C2H6O3]]+	0	1.80	4.77	11.17
174.11217	1.6E+04	0.17	174.11247	-0.3	1.5	C8 H16 O3 N	[[M+H]-[HCOOH]]+	0	1.80	4.77	11.17
184.09897	2.4E+03	0.03	184.09882	0.26	3.5	C9 H14 O3 N	[[M+H]-2[H2O]]+	0	1.80	4.77	11.17
202.10729	1.5E+04	0.16	202.10738	-0.46	2.5	C9 H16 O4 N	[[M+H]-[H2O]]+	0	1.80	4.77	11.17
220.11799	9.69E+06	100	220.11795	0.18	1.5	C9 H18 N O5	[[M+H]]+	0	1.80	4.77	11.17
221.12135	6.84E+05	7.06	221.121305	0.2	1.5	C8 H18 N O5	[[M+H]]+ (13C)	0	1.80	4.77	11.17
222.12166	4.59E+04	0.47	222.122194	-2.4	1.5	C9 H18 N O4 18O	[[M+H]]+ (18O)	0	1.80	4.77	11.17
223.12604	2.26E+03	0.02	223.125549	2.2	1.5	C8 H18 N O4 18O	[[M+H]]+ (13C+18O)	0	1.80	4.77	11.17
238.12923	1.76E+03	0.02	238.128515	3	0.5	C9 H20 N O5	[[M+H]-[H2O]]+	0	1.80	4.77	11.17
242.09992	6.61E+06	68.15	242.099895	0.1	1.5	C9 H17 N Na O5	[[M+Na]]+	0	1.80	4.77	11.17
243.10331	4.44E+05	4.58	243.10325	0.25	1.5	C8 H17 N Na O5	[[M+Na]]+ (13C)	0	1.80	4.77	11.17
244.10363	1.79E+04	0.18	244.104139	-2.09	1.5	C9 H17 N Na O4 18O	[[M+Na]]+ (18O)	0	1.80	4.77	11.17
258.07386	1.29E+05	1.33	258.073833	0.1	1.5	C9 H17 K N O5	[[M+K]]+	0	1.80	4.77	11.17
259.07731	1.49E+03	0.02	259.077188	0.47	1.5	C8 H17 K N O5	[[M+K]]+ (13C)	0	1.80	4.77	11.17
264.082	1.68E+05	1.74	264.08184	0.61	1.5	C9 H16 N Na2 O5	[[M-H+2Na]]+	0	1.80	4.77	11.17
265.08498	3.08E+03	0.03	265.085195	-0.81	1.5	C8 H16 N Na2 O5	[[M-H+2Na]]+ (13C)	0	1.80	4.77	11.17
310.08654	1.10E+03	0.01	310.08732	-2.52	1.5	C10 H18 N Na2 O7	[[M+Na]]+(HCOONa)]+	0	1.80	4.77	11.17
461.21113	6.45E+05	6.65	461.210569	1.22	2.5	C18 H34 N2 Na O10	[[2M+Na]]+	0	1.80	4.77	11.17
463.21439	1.08E+05	1.11	463.213924	0.99	2.5	C17 H34 N2 Na O10	[[2M+Na]]+ (13C)	0	1.80	4.77	11.17
463.2174	9.76E+03	0.1	463.217279	0.26	2.5	C16 H32 N2 Na O10	[[2M+Na]]+ (13C2)	0	1.80	4.77	11.17
483.19256	4.21E+04	0.43	483.192514	0.1	2.5	C18 H33 N2 Na2 O10	[[2M-H+2Na]]+	0	1.80	4.77	11.17

One molecule = one retention time obtained by LC/MS

ESI-mass spectral database
 → Annot. SPI*

* Tools of SPI-LIMS web Interface developed at CEA (CEA/DSV/GIPSI)

Reference compounds analysis : FIA spectrum annotation

SPI-LIMS	
Annotations	Annotation FIA
1. FIA 2. PDBA 3. SPI	Molécule: <input type="text" value="C7 H19 N3"/> Polarité: <input checked="" type="radio"/> Positive <input type="radio"/> Négative Décalage (ppm): <input type="text" value="0"/> Précision (± ppm): <input type="text" value="5"/> Intensité relative ZERO: <input checked="" type="radio"/> OUI <input type="radio"/> NON Fichier MS: <input type="text"/> <input type="button" value="Parcourir..."/> <input type="button" value="Annoter"/>
Peaklists	
Outils	

« Annotation FIA »: to calculate precise m/z of potential ions for a given mass formula and compare them to experimental m/z :

- Isotopes: ^{13}C , ^{34}S , ^{18}O ...
- Adducts: Na, K, Cl...
- Fragments (in source CID)

Reference compounds analysis

Pantothenic acid [M+H]⁺

Retention time

M/Z	Intensity	Relative	M/Z (theo)	Delta (ppm)	RDBE	Composition	Attribution	1.80	4.77	11.17	
72.04431	7.7E+04	0.82	72.04439	-1.11	1.5	C3 H6 O N	[(M+H)-(C6H10O3)-(H2O)]+	1.80	4.77	11.17	
90.05482	3.6E+05	3.78	90.05496	-1.55	0.5	C3 H8 O2 N	[(M+H)-(C6H10O3)]+	1.80	4.77	11.17	
116.034	2.7E+04	0.29	116.03422	-0.22	2.5	C7 H10 O N	[(M+H)-(C2H6O3)-(H2O)]+	1.80	4.77	11.17	
Fragments ions and their isotopes				26	-2.85	2.5	C7 H12 O2 N	[(M+H)-(C2H6O3)]+	1.80	4.77	11.17
				47	-0.3	1.5	C8 H16 O3 N	[(M+H)-(HCOOH)]+	1.80	4.77	11.17
				82	0.26	3.5	C9 H14 O3 N	[(M+H)-2(H2O)]+	1.80	4.77	11.17
				38	-0.46	2.5	C9 H16 O4 N	[(M+H)-(H2O)]+	1.80	4.77	11.17
				Pseudo-molecular ion and its isotopes				95	0.18	1.5	C9 H18 N O5
05	0.2	1.5	C8 13C H18 N O5					[(M+H)]+ (13C)	1.80	4.77	11.17
94	-2.4	1.5	C9 H18 N O4 18O					[(M+H)]+ (18O)	1.80	4.77	11.17
49	2.2	1.5	C8 13C H18 N O4 18O					[(M+H)]+ (13C+18O)	1.80	4.77	11.17
238.12923	1.76E+03	0.02	238.128515	3	0.5	C9 H20 N O6	[(M+H)+(H2O)]+	1.80	4.77	11.17	
242.09992	6.61E+06	68.15	242.099895	0.1	1.5	C9 H17 N Na O5	[(M+Na)]+	1.80	4.77	11.17	
243.10331	4.44E+05	4.58	243.10325	0.25	1.5	C8 13C H17 N Na O5	[(M+Na)]+ (13C)	1.80	4.77	11.17	
244.10363	1.79E+04	0.18	244.104139	-2.09	1.5	C9 H17 N Na O4 18O	[(M+Na)]+ (18O)	1.80	4.77	11.17	
258.07386	1.29E+05	1.33	258.073833	0.1	1.5	C9 H17 K N O5	[(M+K)]+	1.80	4.77	11.17	
259.07731	1.49E+03	0.02	259.077188	0.47	1.5	C8 13C H17 K N O5	[(M+K)]+ (13C)	1.80	4.77	11.17	
264.082	1.68E+05	1.74	264.08184	0.61	1.5	C9 H16 N Na2 O5	[(M-H+2Na)]+	1.80	4.77	11.17	
265.08498	3.08E+03	0.03	265.085195	-0.81	1.5	C8 13C H16 N Na2 O5	[(M-H+2Na)]+ (13C)	1.80	4.77	11.17	
310.08654	1.10E+03	0.01	310.08732	-2.52	1.5	C10 H18 N Na2 O7	[(M+Na)+(HCOONa)]+	1.80	4.77	11.17	
Adducts ions and their isotopes				69	1.22	2.5	C18 H34 N2 Na O10	[(2M+Na)]+	1.80	4.77	11.17
				24	0.99	2.5	C17 13C H34 N2 Na O10	[(2M+Na)]+ (13C)	1.80	4.77	11.17
				79	0.26	2.5	C16 13C2 H34 N2 Na O10	[(2M+Na)]+ (13C2)	1.80	4.77	11.17
				14	0.1	2.5	C18 H33 N2 Na2 O10	[(2M-H+2Na)]+	1.80	4.77	11.17

Reference compounds analysis : Mass spectrum annotation

SPI-LIMS

Annotations	Annotation SPI
<p>1. FIA 2. PDDBA 3. SPI</p>	<p>Polarité: <input checked="" type="radio"/> Positive <input type="radio"/> Négative</p> <p>Décalage (ppm): <input style="width: 100px;" type="text" value="0"/></p> <p>Précision (± ppm): <input style="width: 100px;" type="text" value="5"/></p> <p>Temps de rétention: <input checked="" type="checkbox"/></p> <p>Intervalle de confiance ($x \pm RT^y$): $x =$ <input style="width: 50px;" type="text" value="5"/> $y =$ <input style="width: 50px;" type="text" value="0.80"/></p> <p>Colonnes chromatographiques:</p> <ul style="list-style-type: none"> <input type="checkbox"/> FIA <input type="checkbox"/> HPLC (C18) QTOF <input type="checkbox"/> HSF5 <input type="checkbox"/> UPLC (C18) <input type="checkbox"/> UPLC (C8) <p>Fichier MS: <input style="width: 150px;" type="text"/> <input type="button" value="Parcourir..."/></p> <p style="text-align: center;"><input type="button" value="Annoter"/></p>
Peaklists	
Outils	

Reference compounds analysis: annotation using CEA spectral database

Pantothenic acid [M+H]⁺

M/Z	Intensity	Relative	M/Z (theo)	Delta (ppm)	RDBE	Composition	Attribution	FIA	UPLC (C8)	UPLC (C18)	HSF5
72.04431	7.7E+04	0.82	72.04439	-1.11	1.5	C3 H6 O N	[(M+H)-(C6H10O3)-(H2O)]+	0	1.80	4.77	11.17
90.05482	3.6E+05	3.78	90.05496	-1.55	0.5	C3 H8 O2 N	[(M+H)-(C6H10O3)]+	0	1.80	4.77	11.17
116.034	2.7E+04	0.29	116.03422	-0.22	2.5	C7 H10 O N	[(M+H)-(C2H6O3)-(H2O)]+	0	1.80	4.77	11.17
142.08585	1.0E+03	0.01	142.08626	-2.85	2.5	C7 H12 O2 N	[(M+H)-(C2H6O3)]+	0	1.80	4.77	11.17
174.11217	1.6E+04	0.17	174.11247	-0.3	1.5	C8 H16 O3 N	[(M+H)-(HCOOH)]+	0	1.80	4.77	11.17
184.09687	2.4E+03	0.03	184.09682	0.26	3.5	C9 H14 O3 N	[(M+H)-2(H2O)]+	0	1.80	4.77	11.17
202.10729	1.5E+04	0.16	202.10738	-0.46	2.5	C9 H16 O4 N	[(M+H)-(H2O)]+	0	1.80	4.77	11.17
220.11799	9.69E+06	100	220.11795	0.18	1.5	C9 H18 N O5	[(M+H)]+	0	1.80	4.77	11.17
221.12135	6.84E+05	7.06	221.121305	0.2	1.5	C8 13C H18 N O5	[(M+H)]+ (13C)	0	1.80	4.77	11.17
222.12166	4.59E+04	0.47	222.122194	-2.4	1.5	C9 H18 N O4 18O	[(M+H)]+ (18O)	0	1.80	4.77	11.17
223.12604	2.26E+03	0.02	223.125549	2.2	1.5	C8 13C H18 N O4 18O	[(M+H)]+ (13C+18O)	0	1.80	4.77	11.17
238.12923	1.76E+03	0.02	238.128515	3	0.5	C9 H20 N O6	[(M+H)+(H2O)]+	0	1.80	4.77	11.17
242.09992	6.61E+06	68.15	242.099895	0.1	1.5	C9 H17 N Na O5	[(M+Na)]+	0	1.80	4.77	11.17
243.10331	4.44E+05	4.58	243.10325	0.25	1.5	C8 13C H17 N Na O5	[(M+Na)]+ (13C)	0	1.80	4.77	11.17
244.10363	1.79E+04	0.18	244.104139	-2.09	1.5	C9 H17 N Na O4 18O	[(M+Na)]+ (18O)	0	1.80	4.77	11.17
258.07386	1.29E+05	1.33	258.073833	0.1	1.5	C9 H17 K N O5	[(M+K)]+	0	1.80	4.77	11.17
259.07731	1.49E+03	0.02	259.077188	0.47	1.5	C8 13C H17 K N O5	[(M+K)]+ (13C)	0	1.80	4.77	11.17
264.082	1.68E+05	1.74	264.08184	0.61	1.5	C9 H16 N Na2 O5	[(M-H+2Na)]+	0	1.80	4.77	11.17
265.08498	3.08E+03	0.03	265.085195	-0.81	1.5	C8 13C H16 N Na2 O5	[(M-H+2Na)]+ (13C)	0	1.80	4.77	11.17
310.08654	1.10E+03	0.01	310.08732	-2.52	1.5	C10 H18 N Na2 O7	[(M+Na)+(HCOONa)]+	0	1.80	4.77	11.17
461.21113	6.45E+05	6.65	461.210569	1.22	2.5	C18 H34 N2 Na O10	[(2M+Na)]+	0	1.80	4.77	11.17
462.21438	1.08E+05	1.11	462.213924	0.99	2.5	C17 13C H34 N2 Na O10	[(2M+Na)]+ (13C)	0	1.80	4.77	11.17
463.2174	9.76E+03	0.1	463.217279	0.26	2.5	C16 13C2 H34 N2 Na O10	[(2M+Na)]+ (13C2)	0	1.80	4.77	11.17
483.19256	4.21E+04	0.43	483.192514	0.1	2.5	C18 H33 N2 Na2 O10	[(2M-H+2Na)]+	0	1.80	4.77	11.17



Human urines

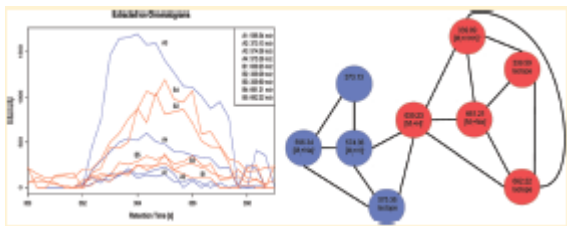
RT	MZ	#	Masse	Composé	Composition	Attribution	UPLC (C18)
4.72	202.107624	1	202.10738	pantothenic acid	C9 H16 O4 N	[(M+H)-(H2O)]+	4.77
4.73	90.054915	1	90.05496	pantothenic acid	C3 H8 O2 N	[(M+H)-(C6H10O3)]+	4.77
4.74	220.117542	1	220.11795	pantothenic acid	C9 H18 N O5	[(M+H)]+	4.77
4.74	221.12168	1	221.121305	pantothenic acid	C8 13C H18 N O5	[(M+H)]+ (13C)	4.77
4.74	222.122088	1	222.122194	pantothenic acid	C9 H18 N O4 18O	[(M+H)]+ (18O)	4.77
4.74	242.100637	1	242.099895	pantothenic acid	C9 H17 N Na O5	[(M+Na)]+	4.77
4.74	258.075314	1	258.073833	pantothenic acid	C9 H17 K N O5	[(M+K)]+	4.77

Tools for annotation and metabolite identification, and data on biofluids begin to be published

analytical
chemistry

CAMERA: An Integrated Strategy for Compound Spectra Extraction and Annotation of Liquid Chromatography/Mass Spectrometry Data Sets

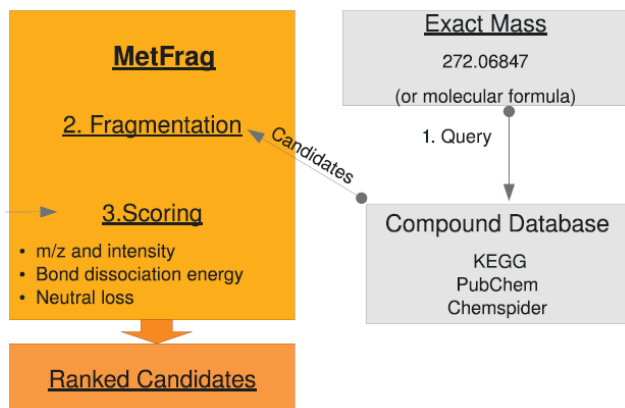
Carsten Kuhl,^{*,†} Ralf Tautenhahn,[‡] Christoph Böttcher,[†] Tony R. Larson,[§] and Steffen Neumann^{*,†}



BMC
Bioinformatics

In silico fragmentation for computer assisted identification of metabolite mass spectra

Sebastian Wolf[†], Stephan Schmidt[†], Matthias Müller-Hannemann[‡], Steffen Neumann[†]



analytical
chemistry

Annotation of the Human Adult Urinary Metabolome and Metabolite Identification Using Ultra High Performance Liquid Chromatography Coupled to a Linear Quadrupole Ion Trap-Orbitrap Mass Spectrometer

Aurelie Roux,[†] Ying Xu,[†] Jean-François Heilier,^{‡,§} Marie-Françoise Olivier,[†] Eric Ezan,[†] Jean-Claude Tabet,[‡] and Christophe Junot^{*,†}

analytical
chemistry

Evaluation of Coupling Reversed Phase, Aqueous Normal Phase, and Hydrophilic Interaction Liquid Chromatography with Orbitrap Mass Spectrometry for Metabolomic Studies of Human Urine

Tong Zhang,^{*,†} Darren J. Creek,^{‡,§} Michael P. Barrett,[‡] Gavin Blackburn,[†] and David G. Watson[†]

Journal of
proteome
research

The Footprints of Gut Microbial–Mammalian Co-Metabolism

Xiaojiao Zheng,^{†,‡} Guoxiang Xie,[†] Aihua Zhao,[†] Linjing Zhao,[†] Chun Yao,[†] Norman H. L. Chiu,[‡] Zhanxiang Zhou,[†] Yuqian Bao,[†] Weiping Jia,^{*,§} Jeremy K. Nicholson,[‡] and Wei Jia^{*,†}

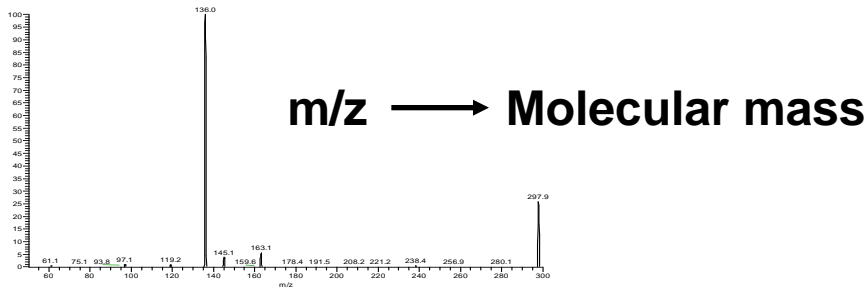
HMDB 3.0—The Human Metabolome Database in 2013

Nucleic Acids Research, 2013, Vol. 41, Database issue D801–D807
doi:10.1093/nar/gks1065

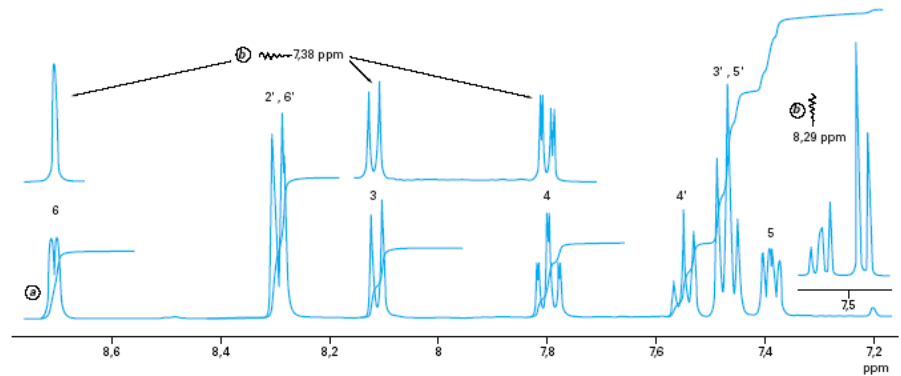
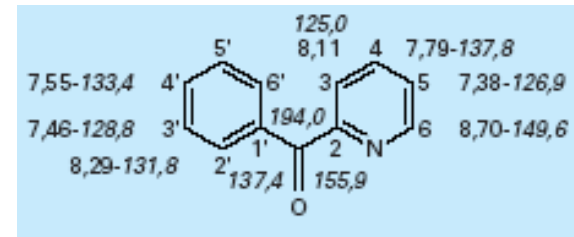
Biochemical data on ~ 40000 metabolites
Spectral data (RMN, MS)

Formal identification of metabolites often requires several complementary analytical tools

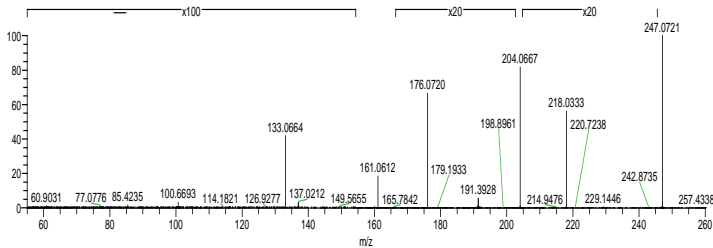
Mass spectrometry



NMR spectroscopy



MS/MS, MSⁿ



Structural information

To discriminate between isomers

L'identification des métabolites:

1. Spectre de masse

- Isoler $[M+H]^+$ ou $[M-H]^-$
- composition élémentaire ($C_xH_yO_z$)
- Isotopes (^{13}C , ^{18}O , $^{34}S...$)

Annotation

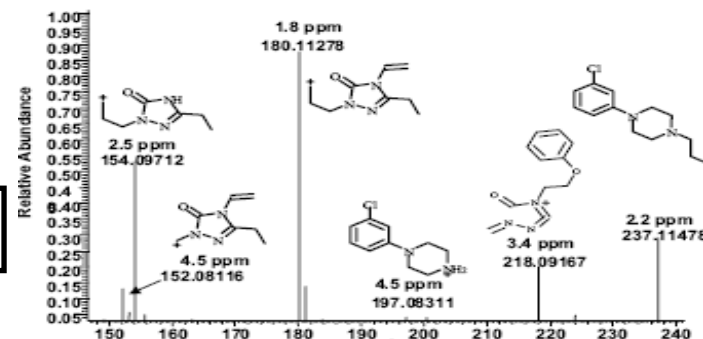
2. Bases de données

3. Spectres de fragmentation (CID)

4. Expériences complémentaires (échange H/D, RMN)

5. Confirmation

- Synthèse chimique
- Analyse LC/MS comparative



Metabolite identification

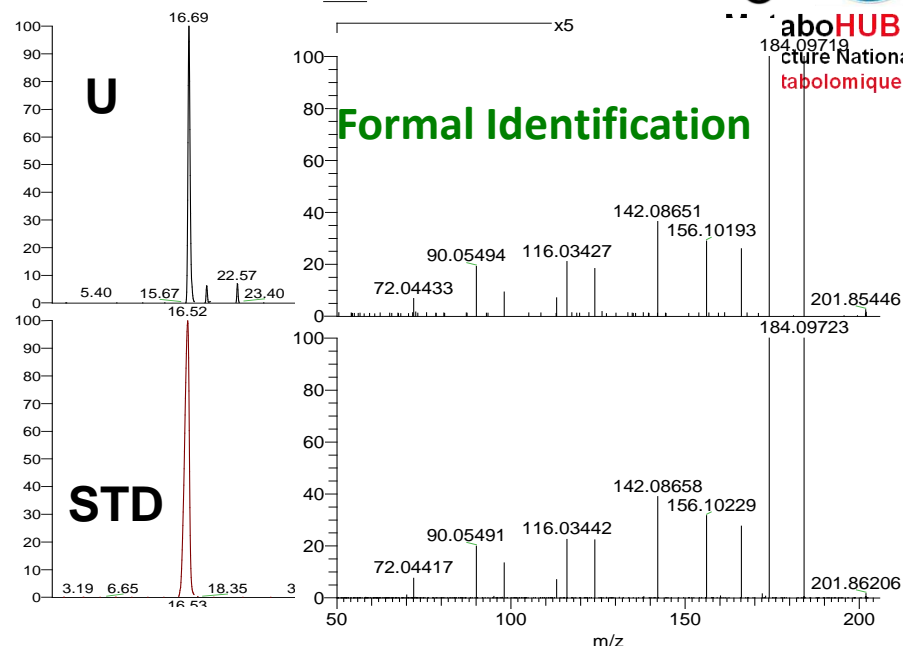
Metabolomics (2007) 3:211–221
DOI 10.1007/s11306-007-0082-2

ORIGINAL ARTICLE

Proposed minimum reporting standards for chemical analysis

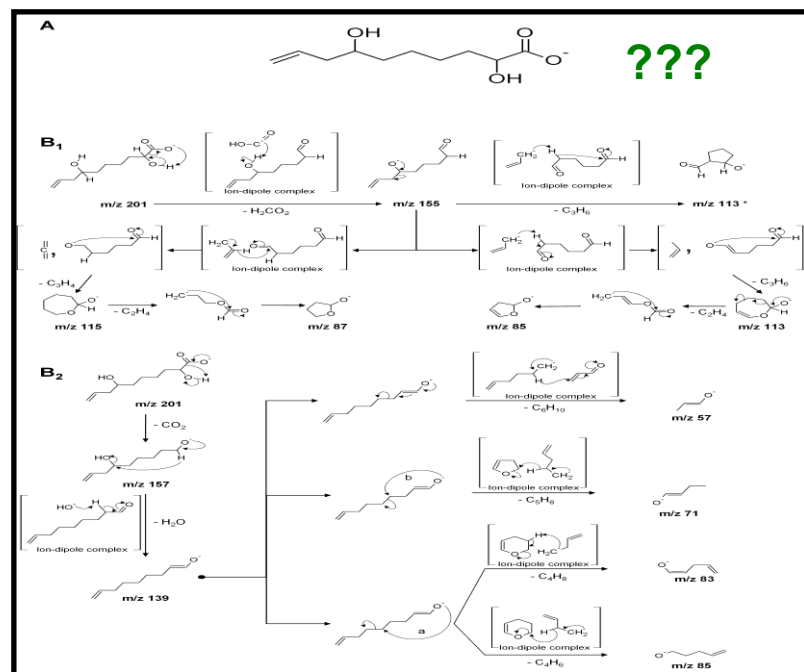
Chemical Analysis Working Group (CAWG) Metabolomics Standards Initiative (MSI)

Lloyd W. Sumner · Alexander Amberg · Dave Barrett · Michael H. Beale · Richard Beger · Clare A. Daykin · Teresa W.-M. Fan · Oliver Fiehn · Royston Goodacre · Julian L. Griffin · Thomas Hankemeier · Nigel Hardy · James Harnly · Richard Higashi · Joachim Kopka · Andrew N. Lane · John C. Lindon · Philip Marriott · Andrew W. Nicholls · Michael D. Reily · John J. Thaden · Mark R. Viant



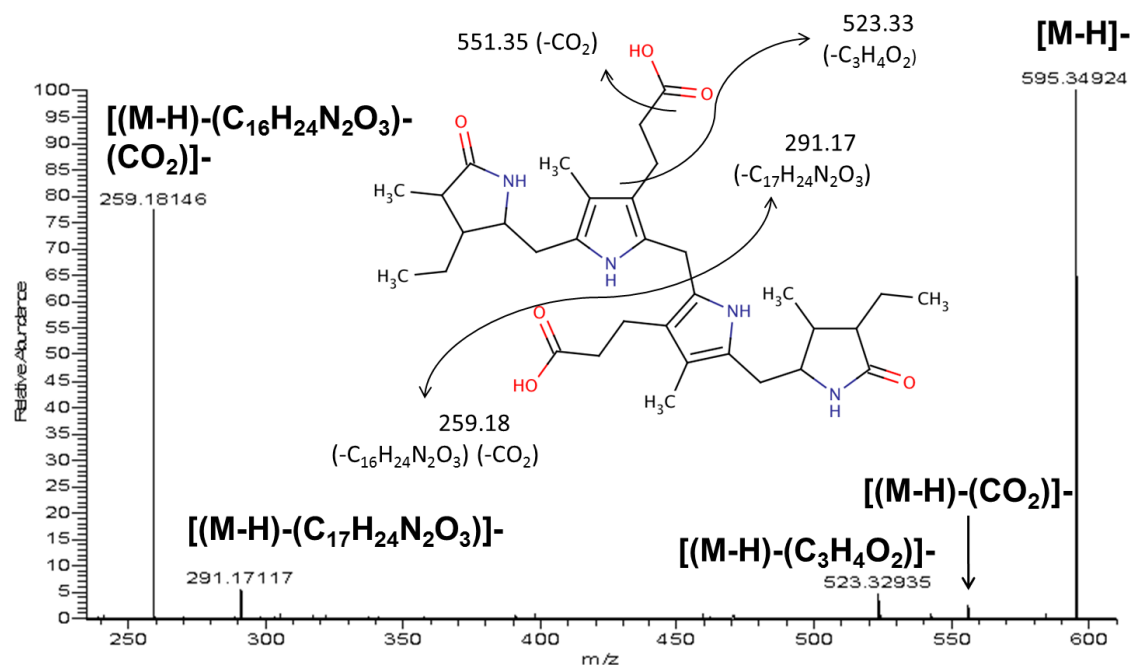
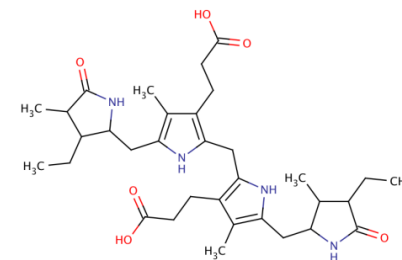
aboHUB
culture Nationale
tabolomique

1. Identified compounds (see below).
2. Putatively annotated compounds (e.g. without chemical reference standards, based upon physicochemical properties and/or spectral similarity with public/commercial spectral libraries).
3. Putatively characterized compound classes (e.g. based upon characteristic physicochemical properties of a chemical class of compounds, or by spectral similarity to known compounds of a chemical class).
4. Unknown compounds—although unidentified or unclassified these metabolites can still be differentiated and quantified based upon spectral data.

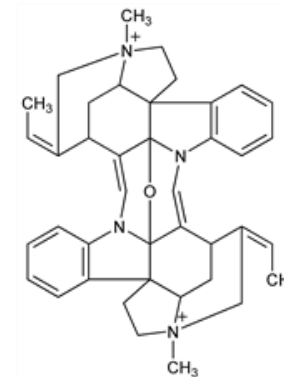


XCMS output			CAMERA output			Inter-sample correlation	Public database annotation
Variable number	m/z	Retention time (min)	isotopes	adduct	pcgroup		
1806	303.1443	9.33	**	**	531	NA	**
4663	593.3312	9.34	[681][M]+	**	512	NA	L-Urobilin
4668	594.3368	9.34	[681][M+1]+	**	512	NA	**
4679	595.3463	9.40	[650][M]+	[M-H]-	394	1.00	C-Curarine / L-Urobilinogen
4682	596.3514	9.40	[650][M+1]+	**	394	0.98	**
4878	631.3256	9.40	**	[M+Cl]-	394	0.96	**
3797	481.2789	9.46	**	**	552	NA	GPCho(10:0/4:0) / GPCho(12:0/2:0)
2763	381.1910	9.53	**	**	627	NA	**
3834	485.1792	9.61	**	**	544	NA	Rutaevin / Nafenopinglucuronide
1255	253.1440	9.67	**	**	556	NA	**

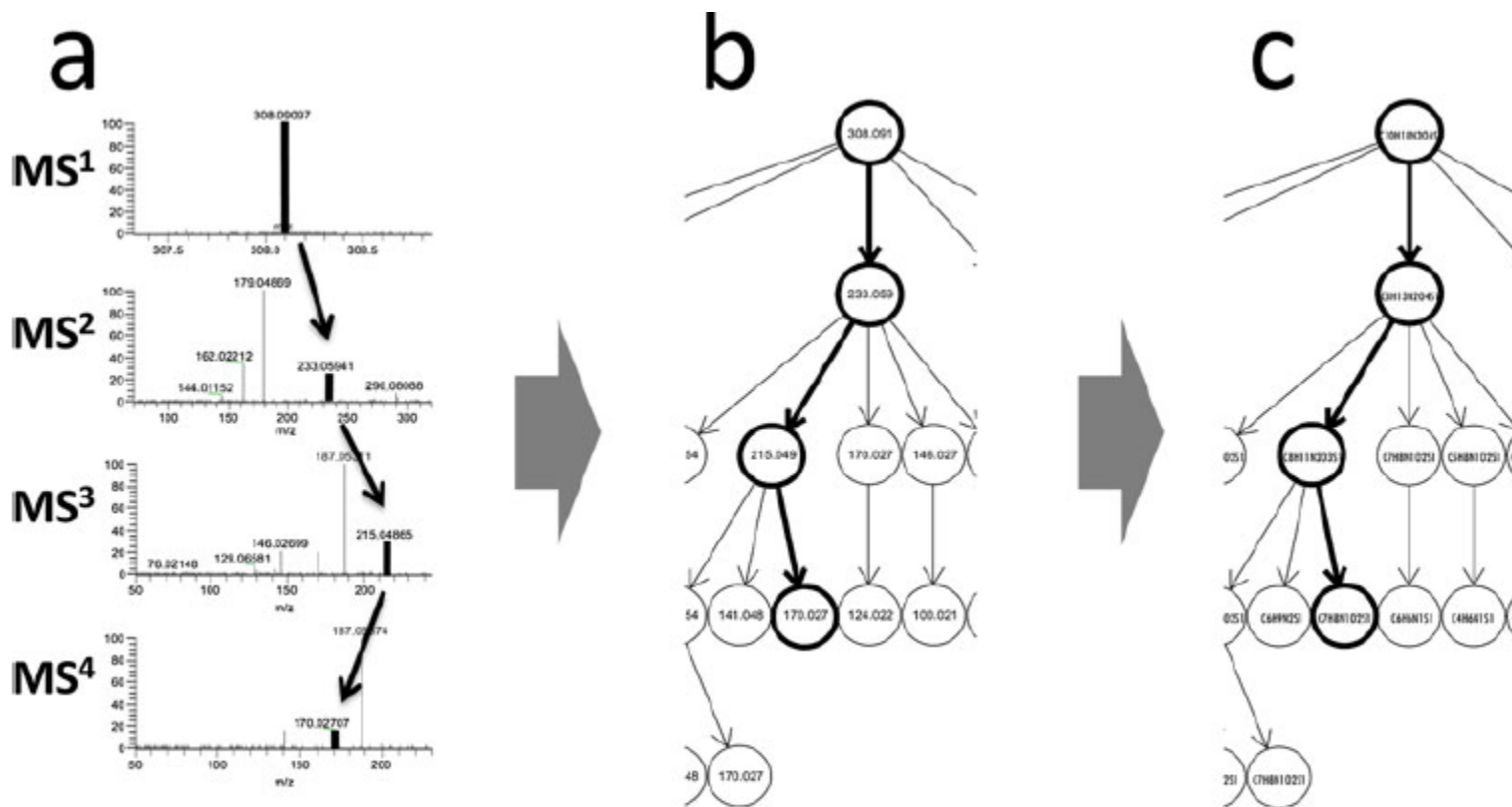
L-Urobilinogen



C-Curarine



Automated analysis of multistage MS (MSⁿ): Spectral trees

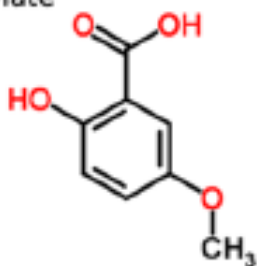


d



Automated analysis of multistage MS (MSⁿ): Spectral trees

Id_7: 5-Methoxysalicylate



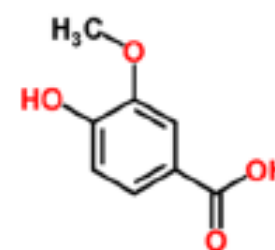
Tree of fragments



Tree of losses



Id_8: Vanillic acid



Tree of fragments

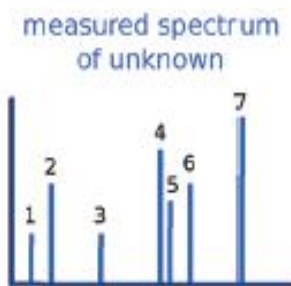


Tree of losses

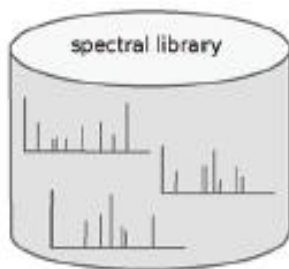
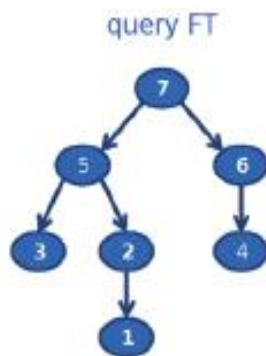


Automated analysis of multistage MS (MSⁿ): Spectral trees

(c) FT-BLAST: searching in reference database



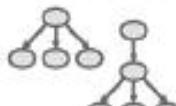
compute FT



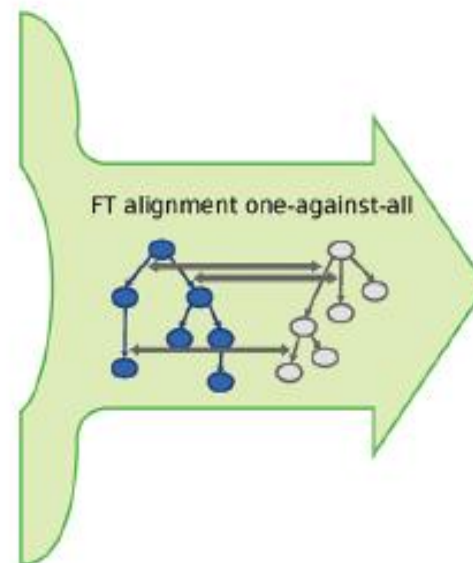
compute FT



compute FT



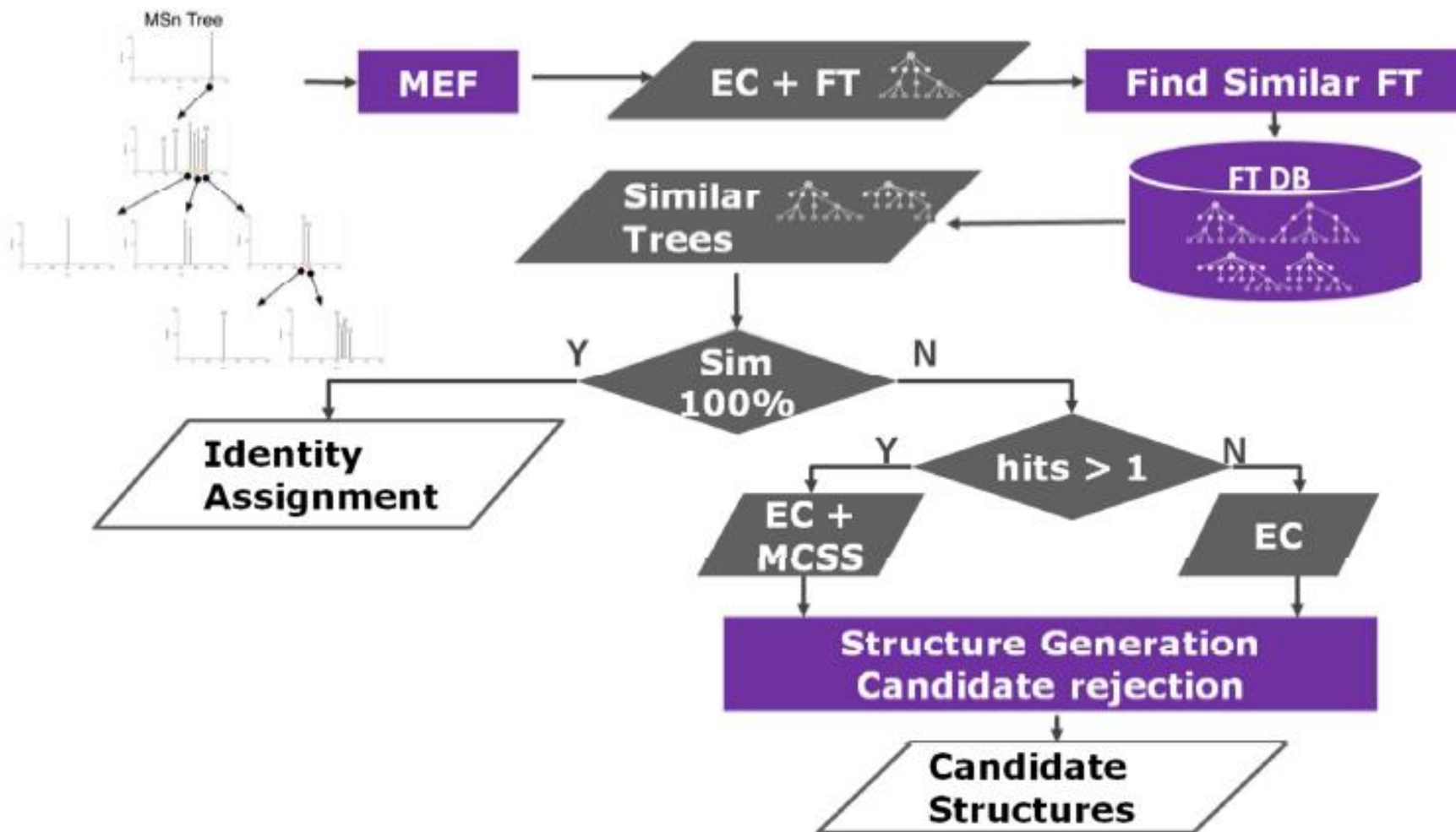
compute FT



search for lactose

hit	q-value
cellobiose	0
trehalose	0
galactose	0
gentiobiose	0
mannose	0
fucose	0
rhamnose	0
DP7	0.17
mannitol	0.17
sorbitol	0.17
delphinidin	0.17
-3-rutinoside	FDR 30%
DP5	0.4
⋮	⋮

Automated analysis of multistage MS (MSⁿ): Spectral trees



*Mise en place d'une méthode d'étalonnage pour
la construction d'une base de données MS/MS en
science métabolomique*


F. Ichou, D. Lesage, C. Junot, J-C. Tabet



Travail actuellement poursuivi et coordonné par **R. Cole**
dans le cadre de l'infrastructure MetaboHUB

Les bases de données

- Majoritairement en ionisation par électron (E.I)
 - ✓ En E.I : beaucoup de fragmentations, absence parfois de l'ion moléculaire M^+ et spectres de masse reproductibles
 - ✓ Nombreuses bases de données et de tailles importantes
 - ✓ Exemple :

	220.000 spectres E.I
---	-------------------------

- En API : Peu de fragmentations, peu reproductible et interférence de la matrice
 - Conséquence : bases de données MS peu fiables
 - Généralement bases de données MS/MS
 - ✓ Problème de reproductibilité en CID à un régime de basse énergie
 - ✓ 2 approches utilisées

Les différentes approches de base de données MS/MS:

1 - Approche non-standardisée

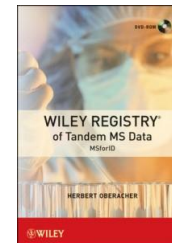
2 - Approche standardisée

Principe reposant sur l'enregistrement de multiples empreintes CID



■ *H.Oberacher et M. Pavlic*⁷

- ✓ Analyses à 10 énergies de collision
- ✓ Elimination de l'ion parent
 - Grande variation de l'abondance
- ✓ Amélioration de l'algorithme de matching



'MSFor ID library'

✓ Autres exemples :



Test



Les différentes approches de bases de données MS/MS:

1 - Approche non-standardisée

2 - Approche standardisée

Etalonnage avec un composé

- ***P. Marquet et al*⁸**

- ✓ Glafenine

- ***C.Hopley and T. Bristow*⁹**

- ✓ Reserpine



Available online at www.sciencedirect.com

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Clinical Biochemistry 38 (2005) 362–372

CLINICAL
BIOCHEMISTRY

Review

LC-MS/MS systematic toxicological analysis: Comparison of MS/MS spectra obtained with different instruments and settings

Robert Jansen, Gérard Lachatre, Pierre Marquet*

Department of Pharmacology-Toxicology, University Hospital, 2 av. Martin-Luther-King, 87042 Limoges cedex, France

Received 18 April 2004; received in revised form 16 July 2004; accepted 5 November 2004
Available online 21 December 2004



RAPID COMMUNICATIONS IN MASS SPECTROMETRY

Rapid Commun. Mass Spectrom. 2008, 22: 1779–1786

Published online in Wiley InterScience (www.interscience.wiley.com) DOI: 10.1002/rcm.3545

Towards a universal product ion mass spectral library – reproducibility of product ion spectra across eleven different mass spectrometers

Chris Hopley¹*, Tony Bristow², Anneke Lubben³, Alec Simpson⁴, Elaine Bull⁴, Katerina Klagkou⁵, Julie Herniman⁶ and John Langley⁶

¹LGC, Queens Road, Teddington TW11 0LY, UK

²AstraZeneca, Macclesfield SK10 2NA, UK

³Department of Chemistry, University of Bath, Bath BA2 7AY, UK

⁴GlaxoSmithKline, Gunnelswood Road, Stevenage SG1 2NY, UK

⁵ThermoFischer, Stafford House, Boundary Way, Hemel Hempstead HP2 7GE, UK

⁶School of Chemistry, University of Southampton, Southampton SO17 1BJ, UK

Received 8 October 2007; Revised 20 March 2008; Accepted 20 March 2008

RCM

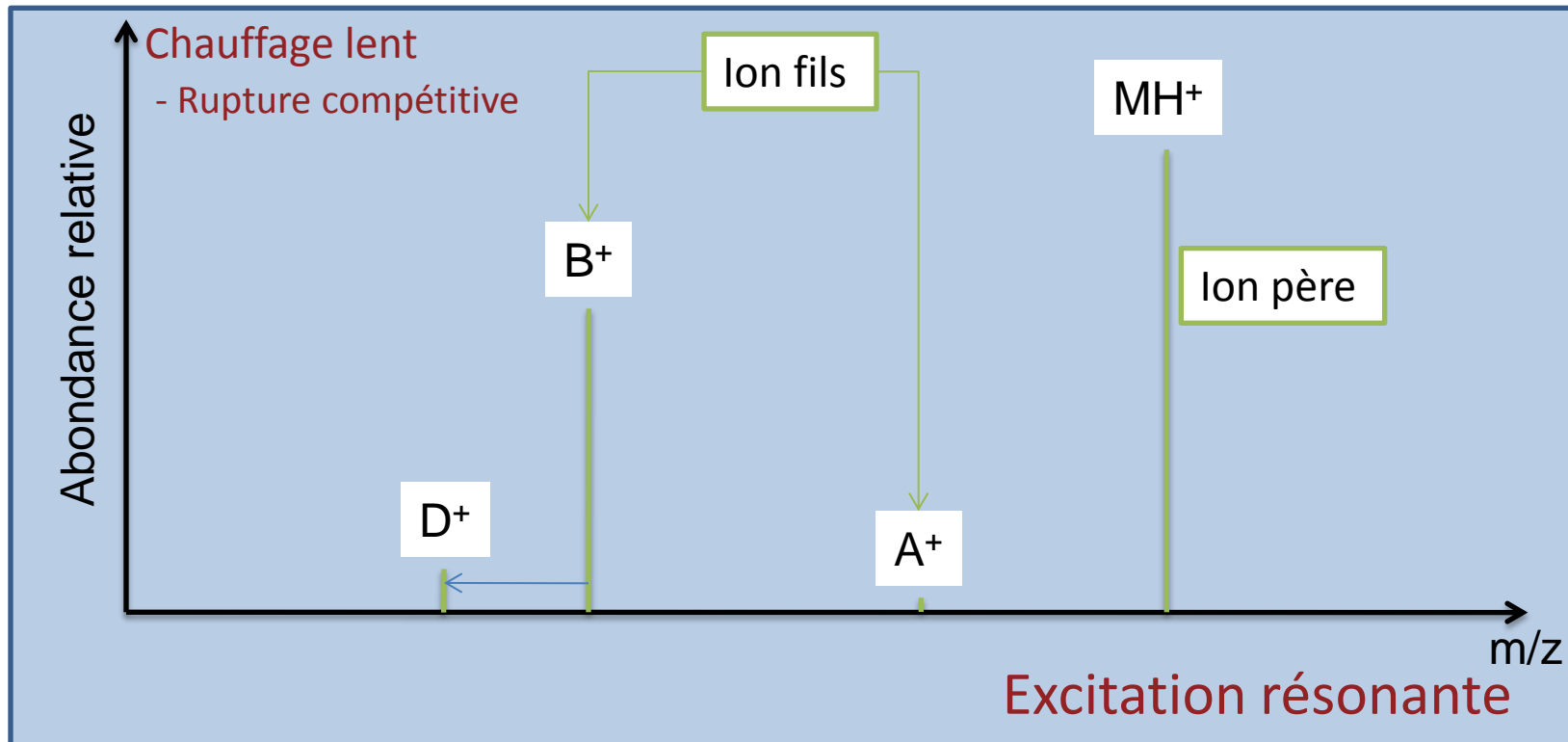


⁸P. Marquet et al, Clin. biochem. 2005, 38, 362

⁹C.Hopley, T. Bristow, Rap. Com. Mass Spectrom. 2008, 22, 1779

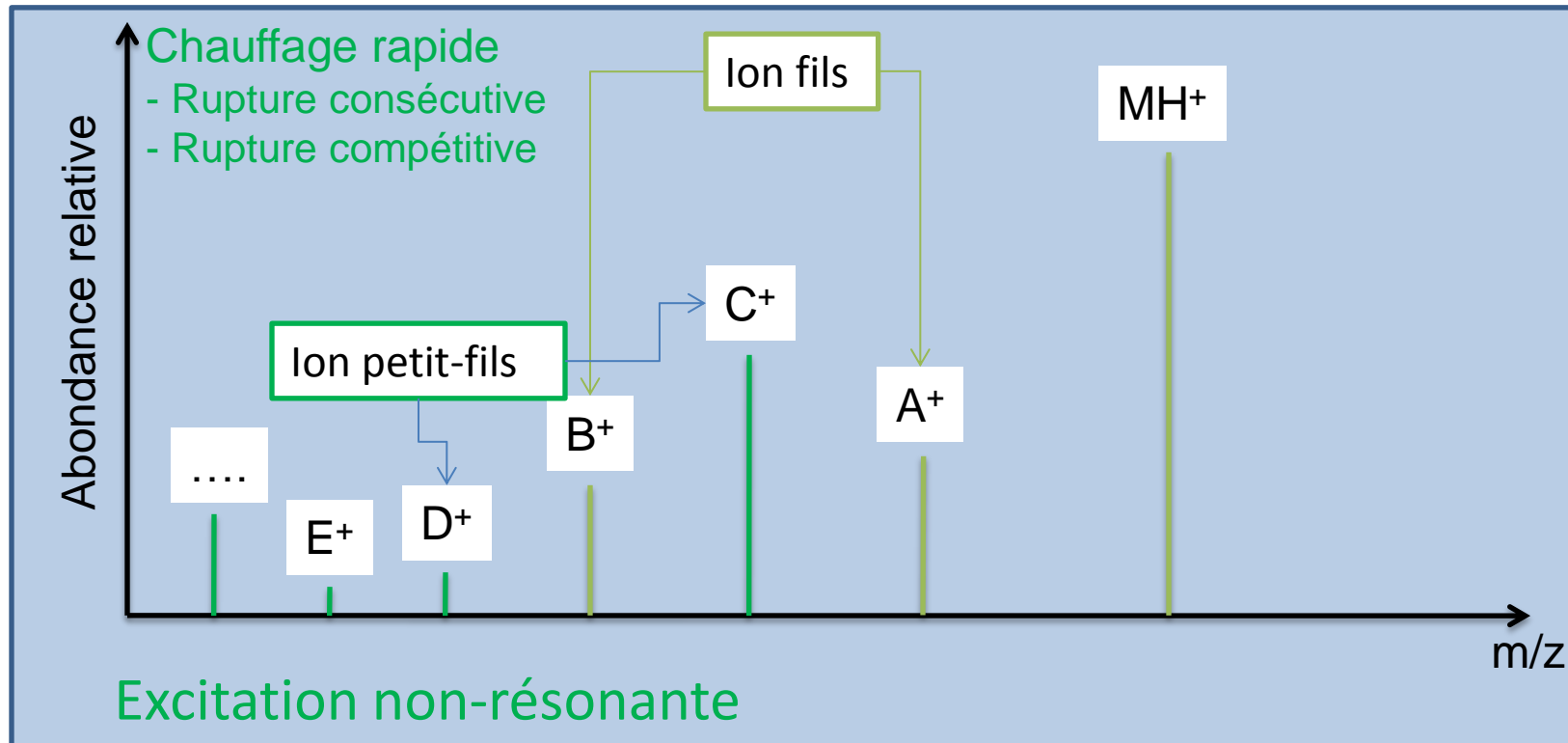
Type d'excitation

Instruments à piégeage d'ions (Piège LTQ, 3D)



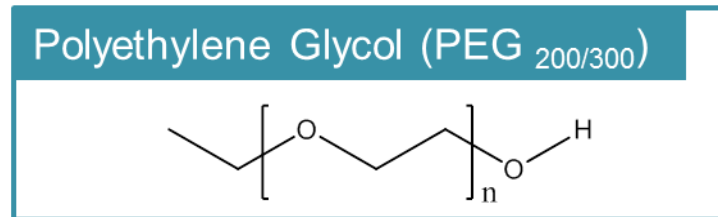
Type d'excitation

Instruments à faisceau d'ions (TQ, QTOF,...)



Procédure d'étalonnage

- Ajuster les ratios des abondances relatives des ions produits par la fragmentation du Polyéthylène glycol (PEG) pour avoir des spectres CID comparables à un spectre de référence



- **Instruments à piégeage d'ions (Résonant) :**
 - ✓ Choisir la même énergie de collision normalisée (15%, 20% et 25%)
 - ✓ Ajuster le temps d'activation
- **Instruments à faisceau d'ions (Non-résonant) :**
 - ✓ Choisir trois énergies de collision
 - ✓ Ajuster la pression de la cellule de collision

Objectifs

Evaluation de la procédure :

Reproductibilité inter-laboratoire sur le même type d'instruments

✓ Est-il possible d'utiliser des spectres de référence enregistrés dans d'autres laboratoires pour identifier les métabolites ?

Comparaison inter-instrumentale

✓ Les spectres CID provenant de différents instruments donnent-ils les mêmes informations ?

Quelles sont les limites de la méthode d'étalonnage ?

✓ Doit-on construire une base de données MS/MS spécifique à chaque type d'instruments (tandems à faisceau d'ions ou à piégeage d'ions)?

Validation de la procédure

- 19 composés :
 - ✓ Panel représentatif de la diversité des problèmes rencontrés en métabolomique
- 9 laboratoires partenaires
- 13 instruments (QTOF, LTQ, Orbitrap)
- Mise en situation avec un logiciel commercial (SmileMS)

Panel de composés

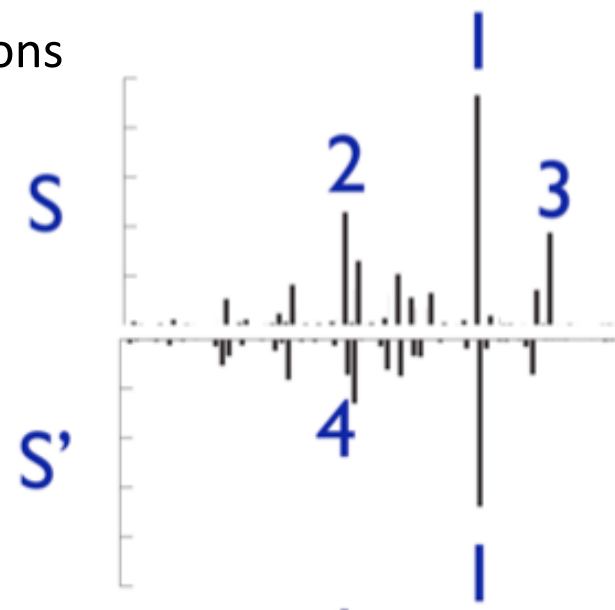
Composé	Masse molaire (g/mol)	Polarité	Caractéristique des composés
Caféine	194	Pos	Groupe A. Peu de voies de dissociations compétitives (< 2 ions fragments dans les pièges à ions)
Myricetine	318	Pos&Neg	
Xanthosine	284	Pos&Neg	
Inosine	268	Pos&Neg	
5-Methoxyindoleacetate	205	Pos	
Acide Indoleacrylique	187	Neg	Groupe B. 2-3 ions fragments
Acide 12-hydroxydodecanoïque	216	Neg	
Acide Xanthurenique	205	Neg	
Dimethoate	229	Pos	
Naringénine	272	Pos	
Tyrosine	181	Pos&Neg	Groupe C. Beaucoup de voies de dissociations (> 3 ions fragments)
Panthenol	205	Pos&Neg	
Acide Cis-aconitique	174	Neg	
Acide Trans-aconitique	174	Neg	
Cystathionine	222	Pos&Neg	
Acetamidrid	222	Pos&Neg	
Arginine	174	Pos&Neg	
Glutathion	307	Pos&Neg	
Carnosine	226	Pos&Neg	

Logiciel SmileMS

➤ Originalité de l'algorithme X-Rank

- Comparaison par rapport aux m/z et indirectement aux abondances relatives des ions
- Classification des fragments par ordre d'intensité des pics
- Compatibilité Inter-instrumentale
 - Extraction par protéowizards des différents formats constructeurs

Conversion sous .mzXML, .mgf



Principe de l'expérience

Comparaison :

- ✓ Inter-QTOF
- ✓ Inter-LTQ
- ✓ Inter-Orbitrap
- ✓ Inter-instrumentale

Deux principaux résultats :

■ Score d'identification

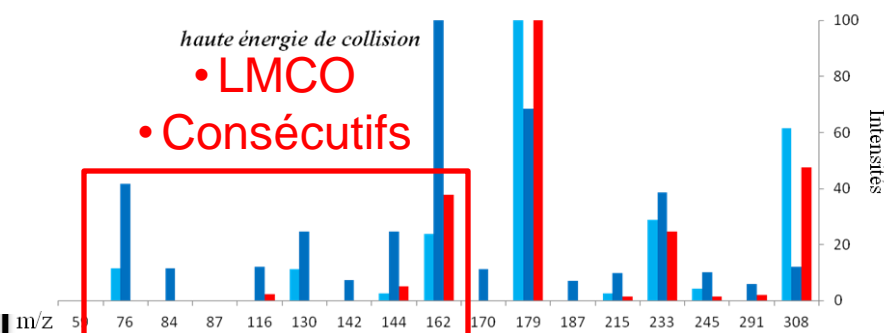
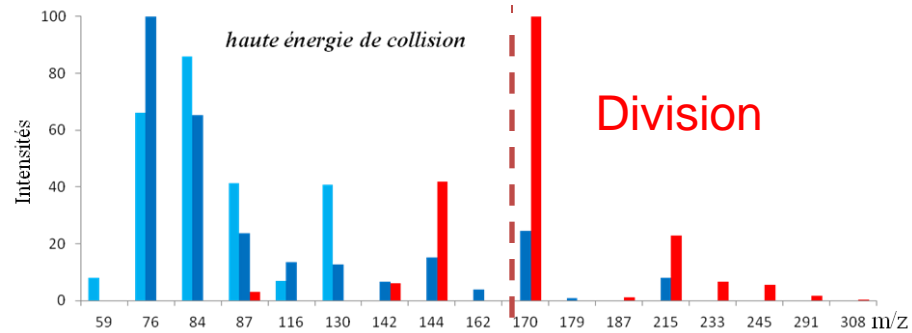
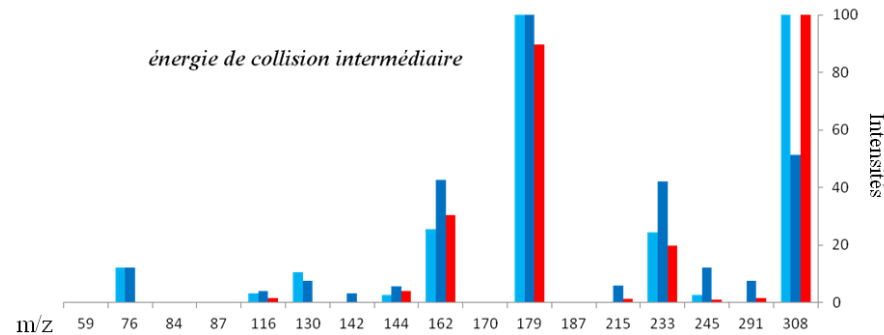
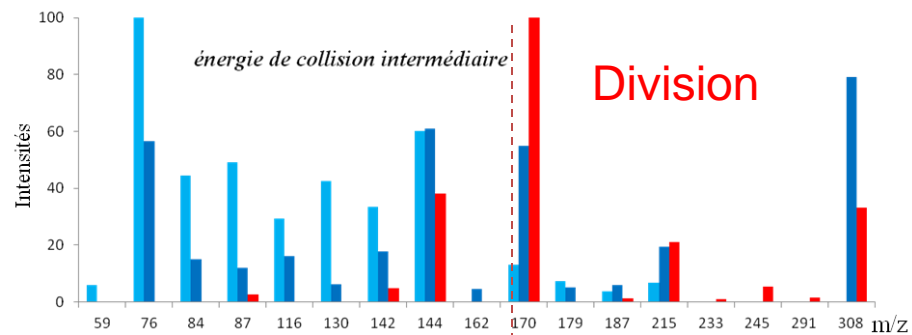
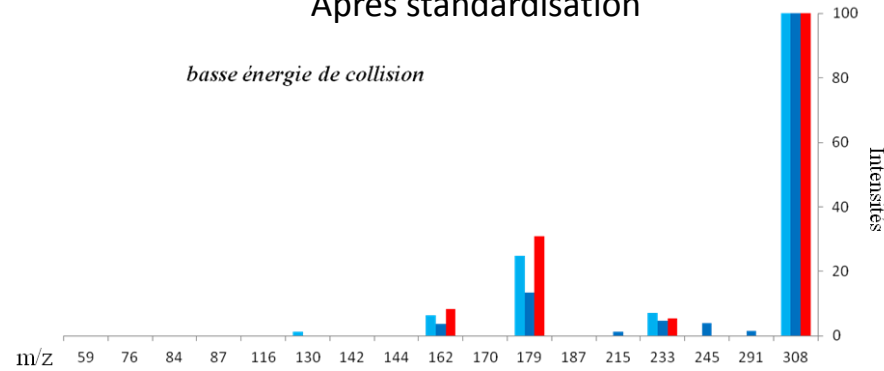
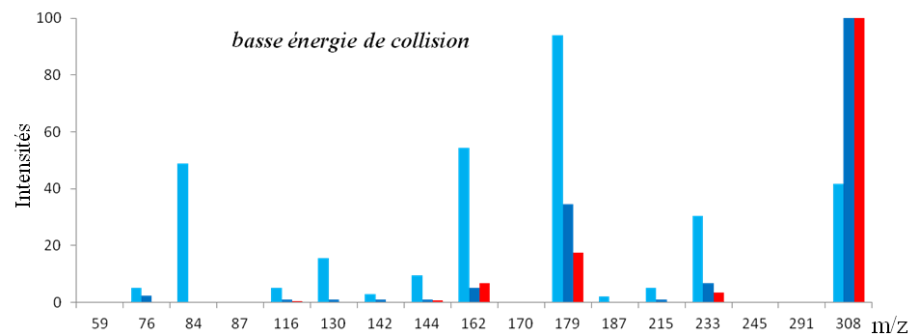
- ✓ Score dépendant de la quantité d'informations contenues dans le spectre CID
- ✓ Nette baisse du score en négatif sauf pour les molécules ayant beaucoup d'ions fragments
 - Mode négatif plus pauvre en fragments

■ Ecart-type

Résultats inter-instrumentaux

Avant standardisation

Après standardisation

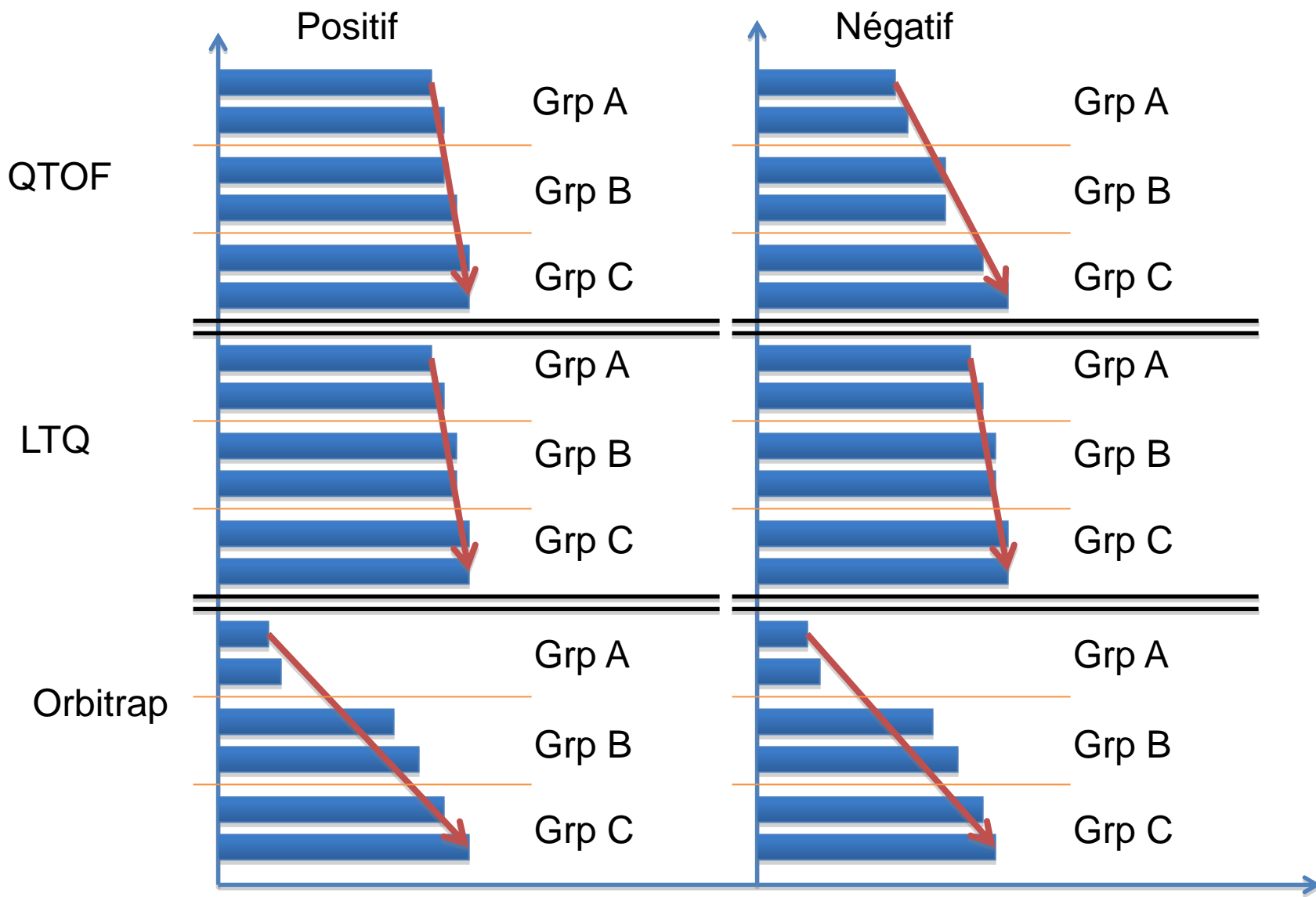


Orbitrap

TQ

QTOF

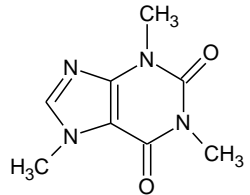
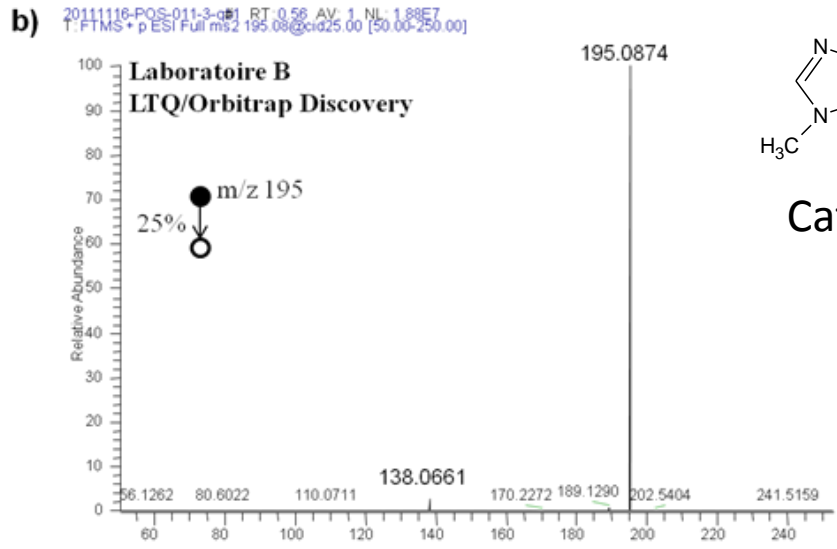
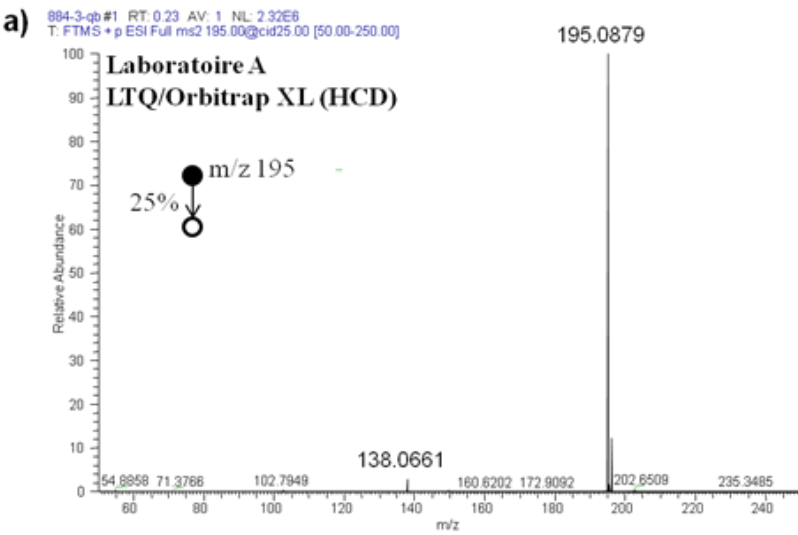
Quantité d'informations



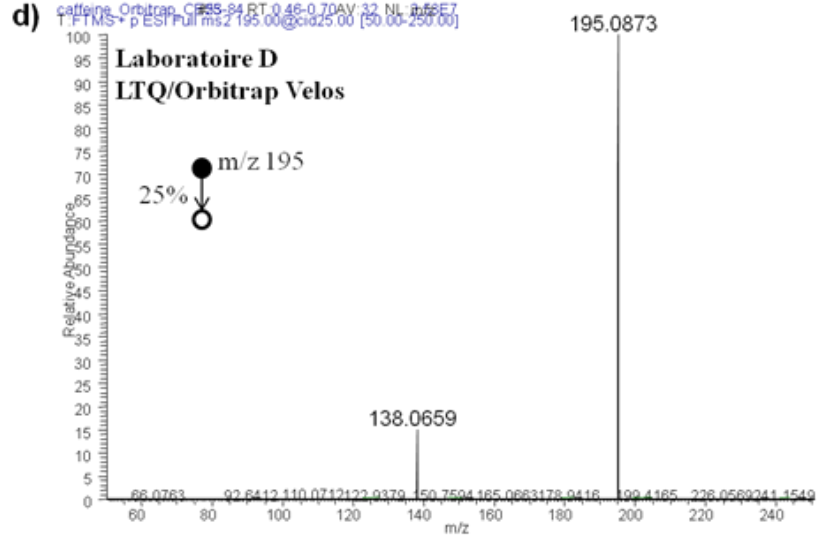
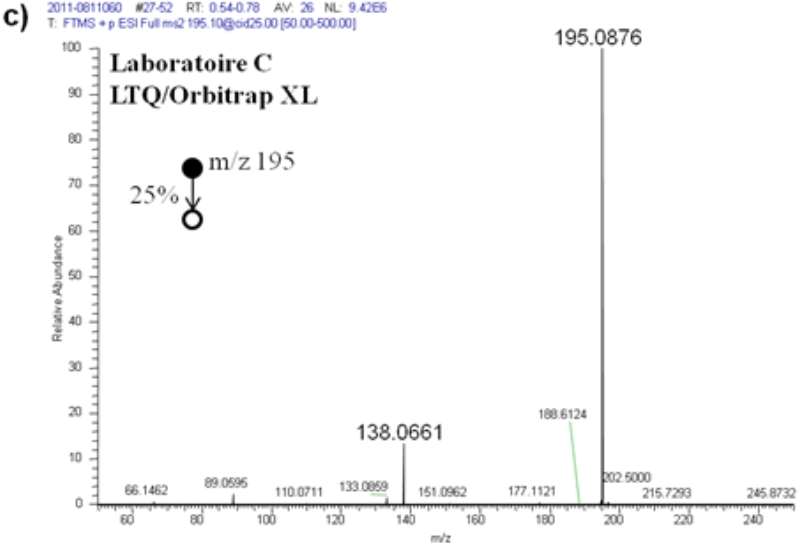
Résultats inter-Orbitrap

Score moyen d'identification et écart-types proches de 0

En mode positif



Caféine



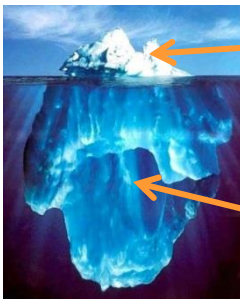
Conclusion

➤ *Procédure de standardisation*

- Meilleure reproductibilité inter-laboratoires et inter-instrumentale
- Intérêts de la procédure d'étalonnage
 - ✓ Contrôle de l'énergie interne
 - ✓ Réduction des ruptures consécutives pour les tandems à faisceau d'ions
 - ✓ Chaque instrument peut être une référence
- 2 Sources de déviation dans les scores
 - ✓ Les molécules avec peu de fragmentations
 - ✓ Les tandems à faisceau d'ions (contrôle énergétique plus difficile)

➤ *Perspectives*

- ✓ Validation et construction de la librairie
- ✓ Projet MétaboHUB



Standards commerciaux

Composés issus de matrices biologiques,...

Several methods have to be implemented in order to achieve an optimal metabolome coverage

[dx.doi.org/10.1021/ac2030738](https://doi.org/10.1021/ac2030738) | *Anal. Chem.* 2012, 84, 1994–2001

Evaluation of Coupling Reversed Phase, Aqueous Normal Phase, and Hydrophilic Interaction Liquid Chromatography with Orbitrap Mass Spectrometry for Metabolomic Studies of Human Urine

Tong Zhang^{*,†} Darren J. Creek^{‡,§} Michael P. Barrett[‡] Gavin Blackburn[†] and David G. Watson[†]

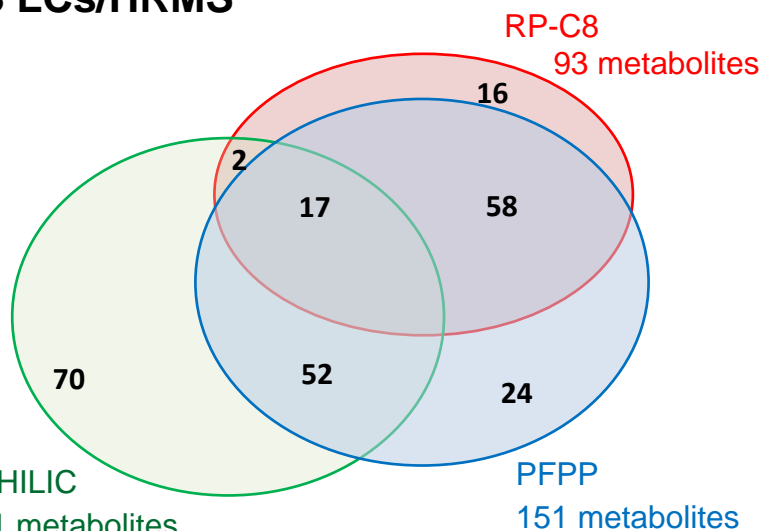
[dx.doi.org/10.1021/ac300586m](https://doi.org/10.1021/ac300586m) | *Anal. Chem.* 2012, 84, 6963–6972

Systematic Evaluation of Extraction Methods for Multiplatform-Based Metabotyping: Application to the *Fasciola hepatica* Metabolome

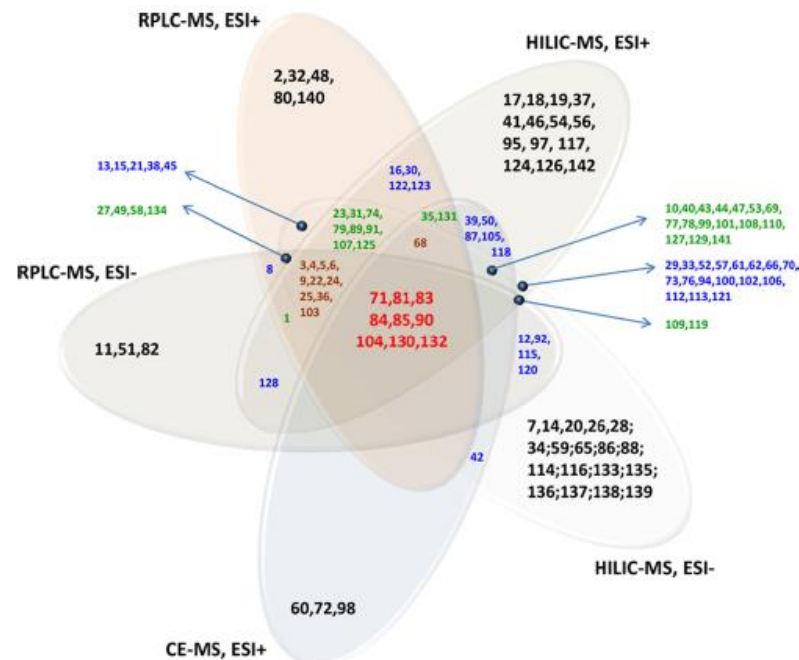
Jasmina Saric[†] Elizabeth J. Want[†] Urs Duthaler[‡] Matthew Lewis[†] Jennifer Keiser[‡] John P. Shockcor[‡] Gordon A. Ross[#] Jeremy K. Nicholson[†] Elaine Holmes^{*,†} and Marina F. M. Tavares^{*,†,||}

270 metabolites identified in human plasma

3 LCs/HRMS



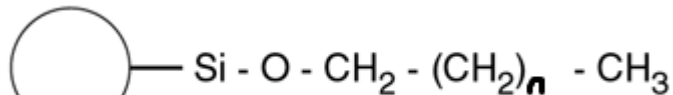
(Boudah et al., *J. Chromatogr. B*, 2014)



The is a need to add other separative dimensions to HRMS

Reverse Phase Liquid Chromatography

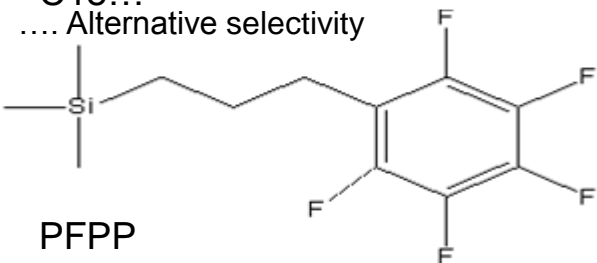
Classic



Alkyl bound silica

C8,
C18...

.... Alternative selectivity



PFP

PentaFluoroPhenylPropyl bound silica

Retention mechanism :

- **Hydrophobic interactions**

Bile acids, Steroid hormones...

Retention mechanism :

- **π - π interactions** for phenyl-based compounds

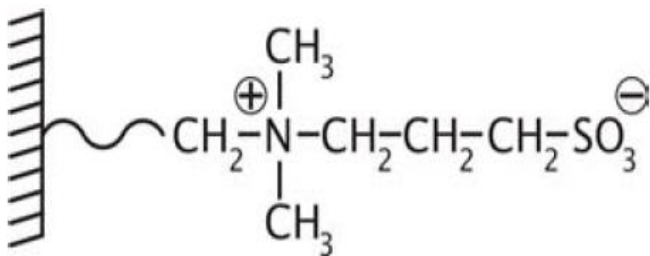
- **dipole-dipole interactions** for halo/polar compounds

Amino acids, cyclic compounds...

Most widely used columns:

- aqueous and organic solvents compatibility
- robustness
- well defined retention mechanisms
- continuous manufacturer improvement

Hydrophilic Interaction Liquid Chromatography



ZIC HILIC

Zwitterionic sulfobetaine bound silica

Retention mechanism :

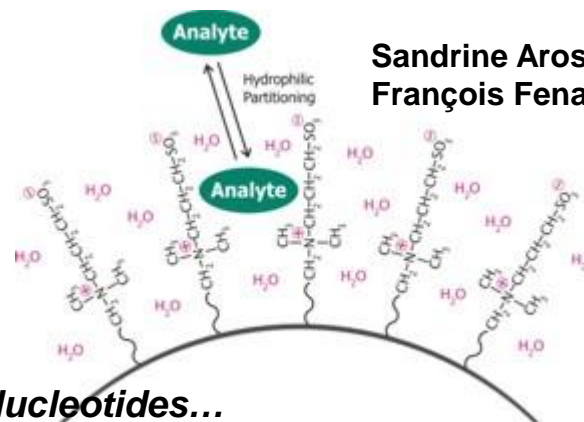
- **hydrophilic partitioning**

from the eluent to the enriched-water layer

- **electrostatic interactions**

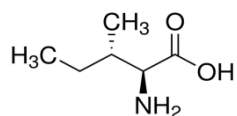
with either positive and negative charges

Carboxylic acids, Sugars, Nucleotides...

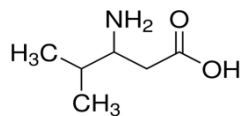


Sandrine Aros-Calt
François Fenaille

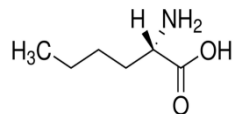
To discriminate between
isomer species



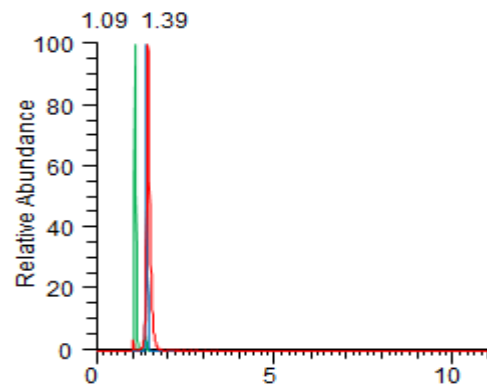
Isoleucine



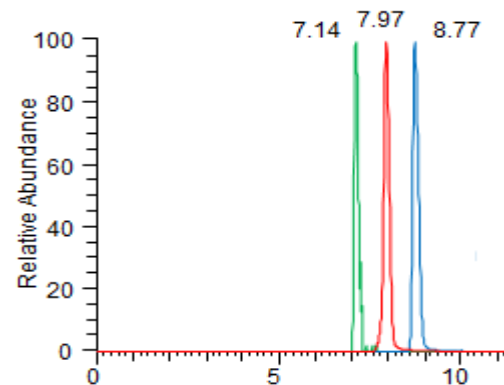
β Leucine



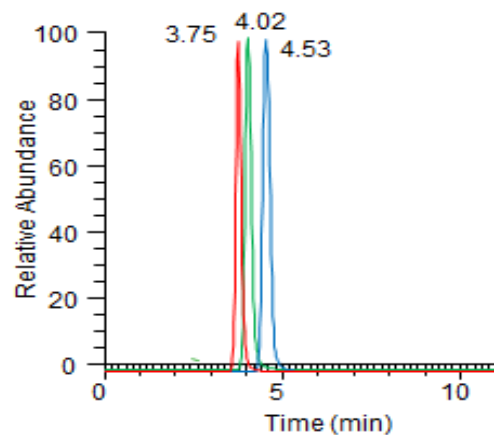
Norleucine



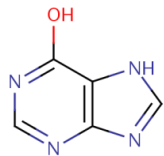
RP-C8



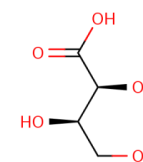
PFPP



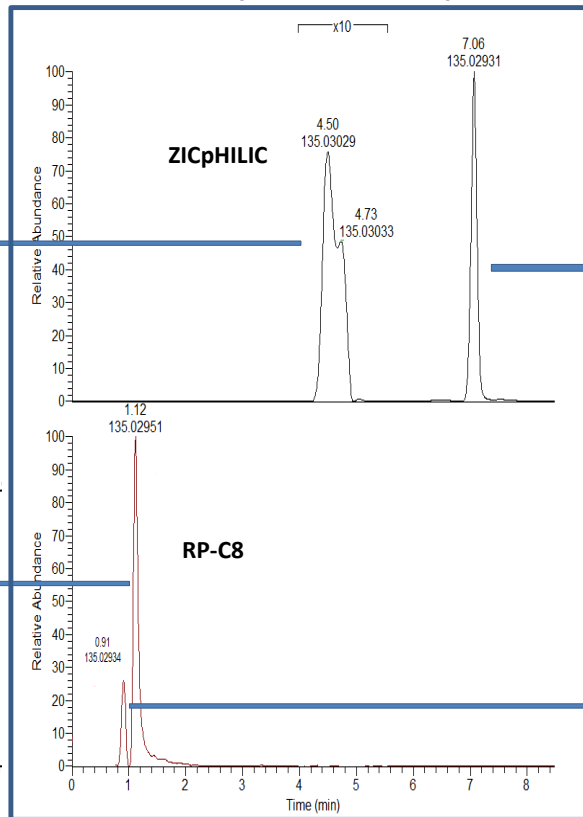
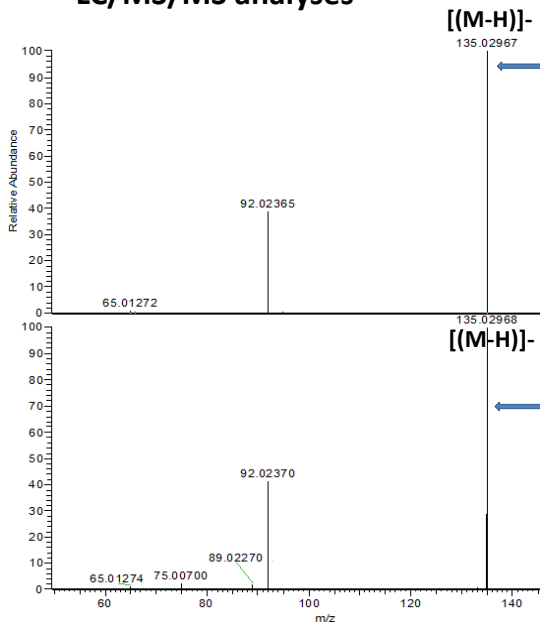
ZICpHILIC



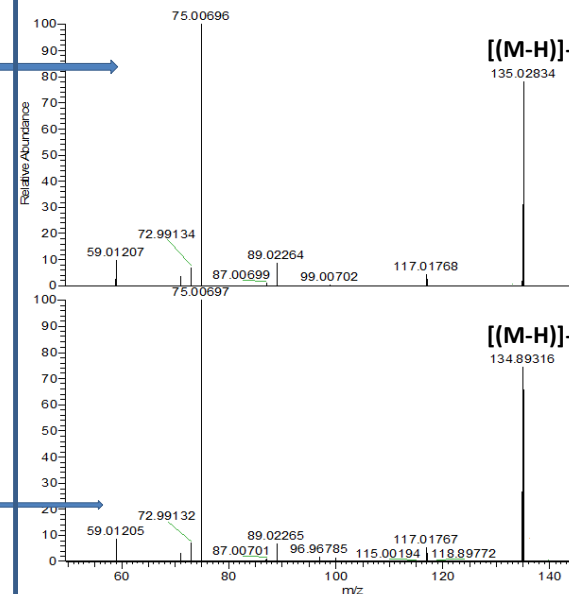
LC/MS analyses of human plasma



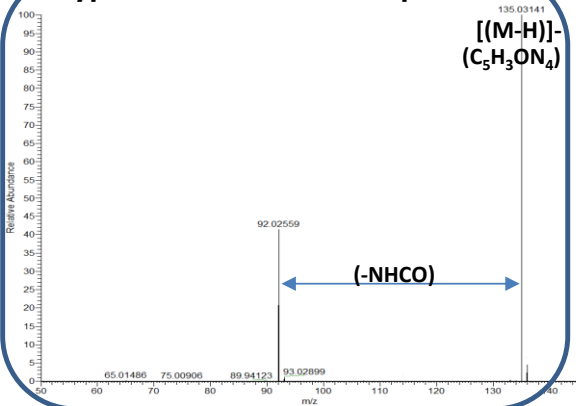
LC/MS/MS analyses



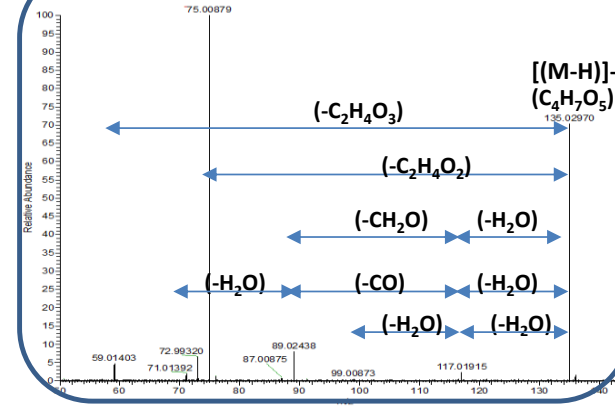
LC/MS/MS analyses



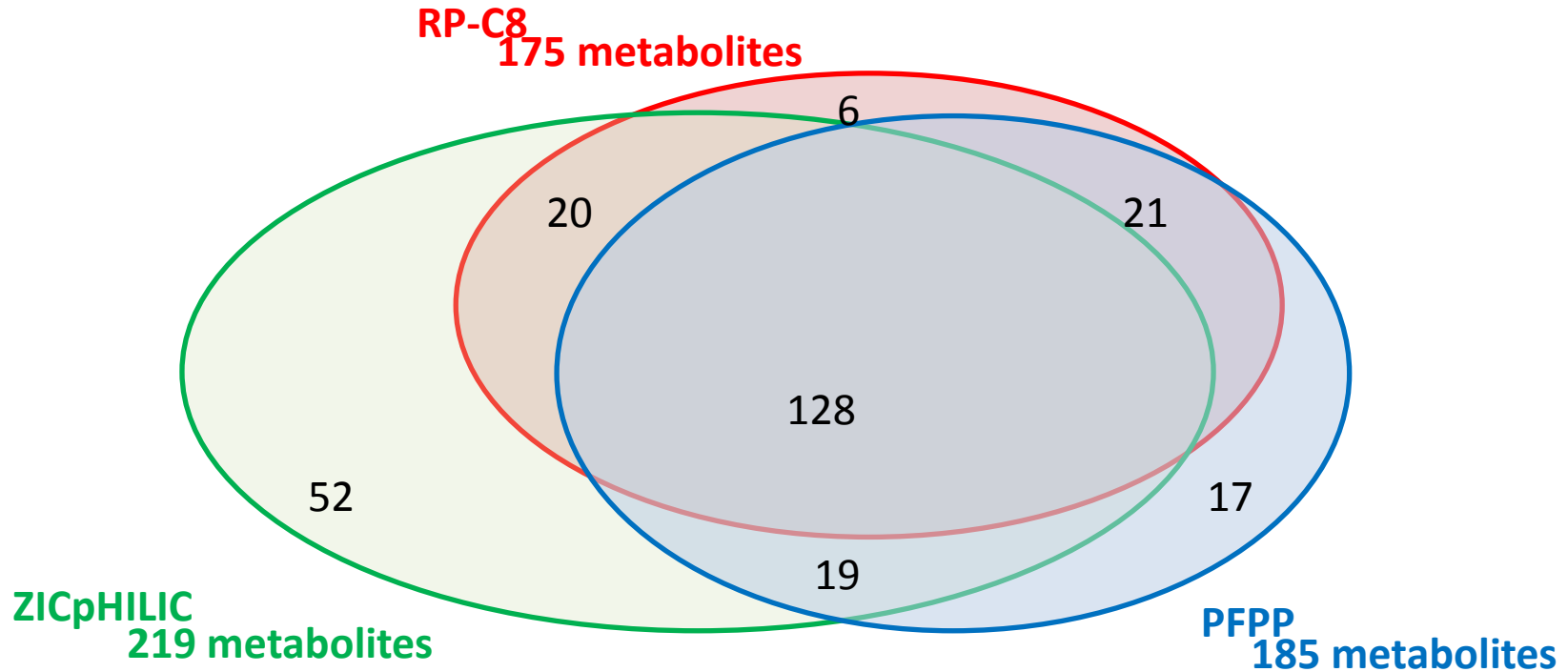
Hypoxanthine reference spectra



Threonic acid reference spectra



Annotation of the human serum metabolome using LC/HRMS



-**266** metabolites were distributed over **209** distinct accurate masses

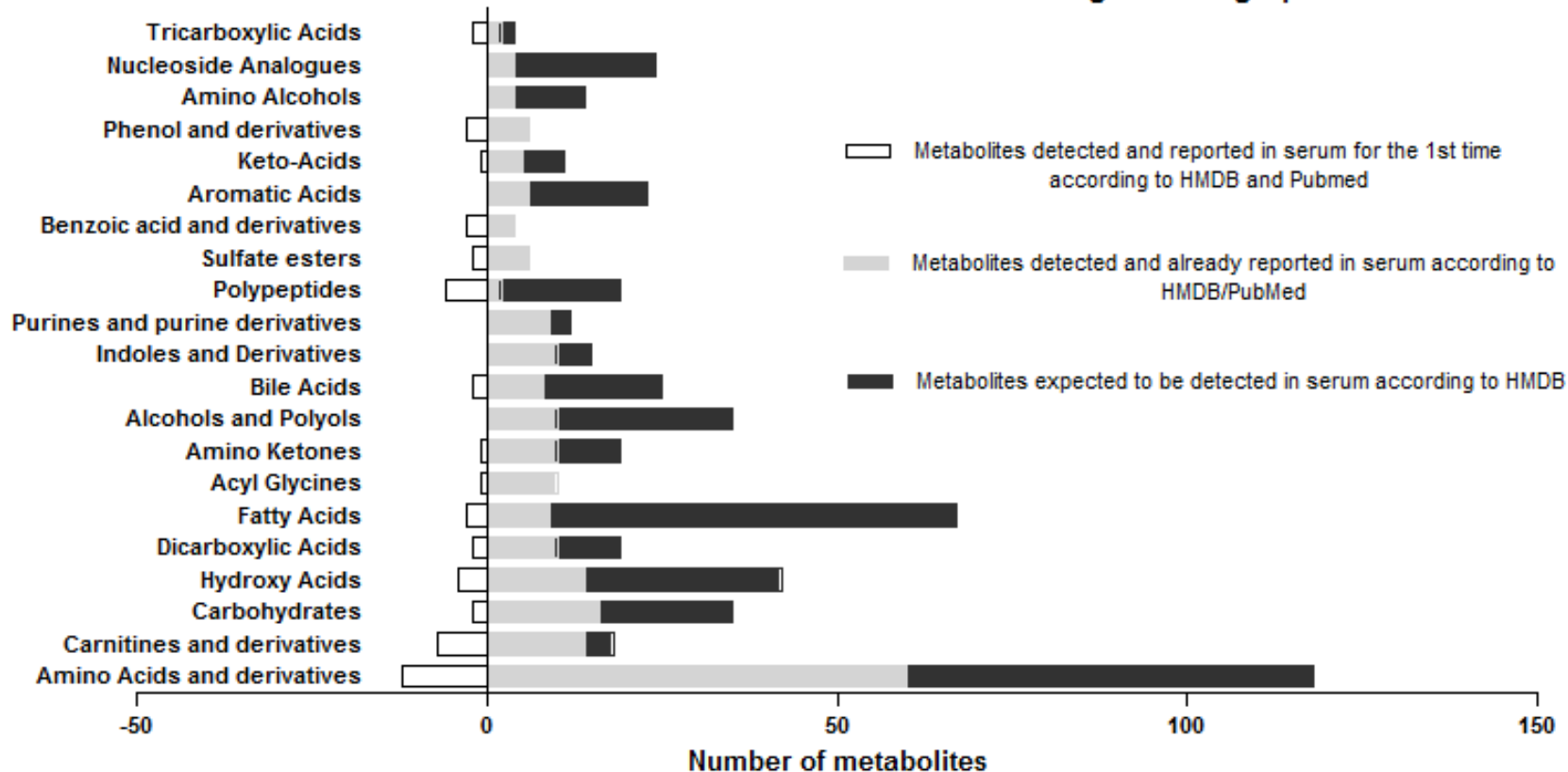
-ESI(-): HILIC extended metabolome coverage 50% and the number of retained metabolites ($k>1$) 75%.

-ESI(+): PFPP conditions improved the retention up to almost 200% of metabolites detected.

-using HILIC (ESI-) and PFPP (ESI+) ensure the detection of 243 out of 266 metabolites in serum samples.

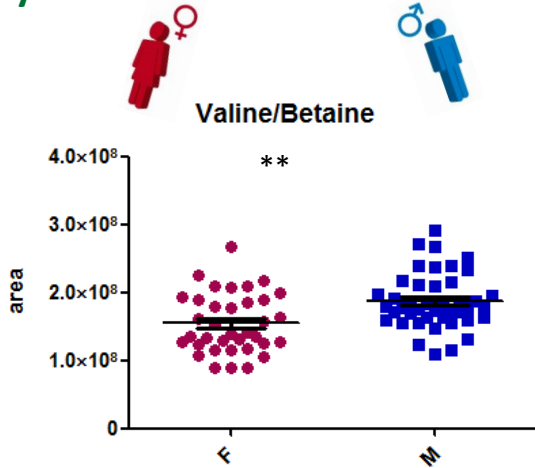
Annotation of the human serum metabolome using LC/HRMS

Distribution of 239 detected serum metabolites* according to bibliographic data

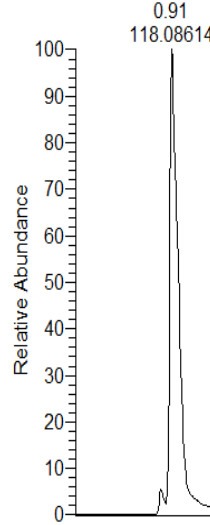


Multiplatform strategy: toward a comprehensive assessment of metabolomic profiles

Alkyl RP vs PFPP



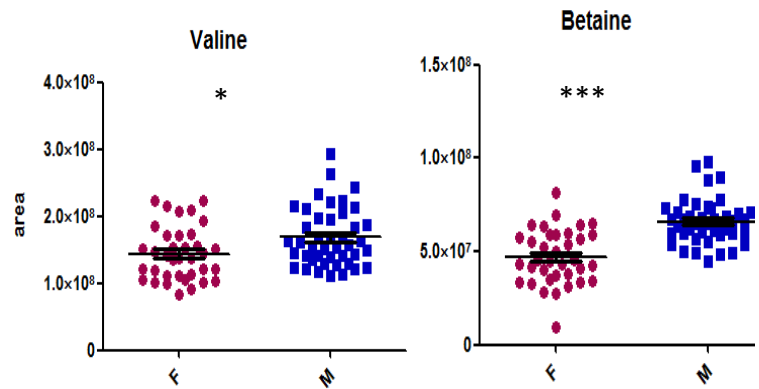
RT: 0.00 - 8.04 SM: 7G



Valine ?
Betaine ?
Both ?

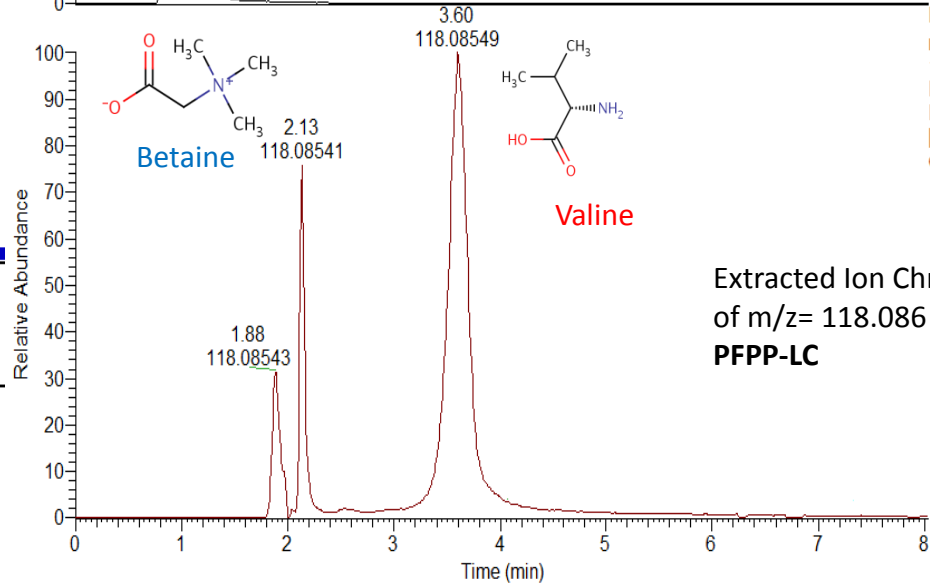
NL: 3.36E7
m/z=
118.08522-118.08710
F: FTMS {1;1} +p ESI
Full ms
[75.00-1000.00] MS
QC-Dil1_Pos_7

Extracted Ion Chromatogram
of m/z= 118.086 obtained in
RP-LC



Proteinogenic
amino acids

Detoxification
reaction : liver,
kidney



NL: 1.17E7
m/z=
118.08522-118.08710
F: FTMS {1;1} +p ESI
Full ms
[75.00-1000.00] MS
qc-dil1_pos_6

Extracted Ion Chromatogram
of m/z= 118.086 obtained in
PFPP-LC

Toward databases of metabolic profiles: it is required to control analytical biases through design of experiment

- Randomization is required
- Batches of ~ 100 injections
- Blank and QC samples

Some «reference» protocols are available:

Human plasma: Dunn WB, Nat. Protocols, 2011

Human urines: Want E., Nat. Protocols, 2010

Pool d'échantillons bio préparés à 3 dilutions et injectés en n=3 en vue du traitement des données

QC-pool + EI_dil-8
QC-pool + EI_dil-8
QC-pool + EI_dil-8

QC-pool + EI_dil-4
QC-pool + EI_dil-4
QC-pool + EI_dil-4

QC-pool + EI_dil-2
QC-pool + EI_dil-2
QC-pool + EI_dil-2

blanc
QC-pool
QC-pool + EI

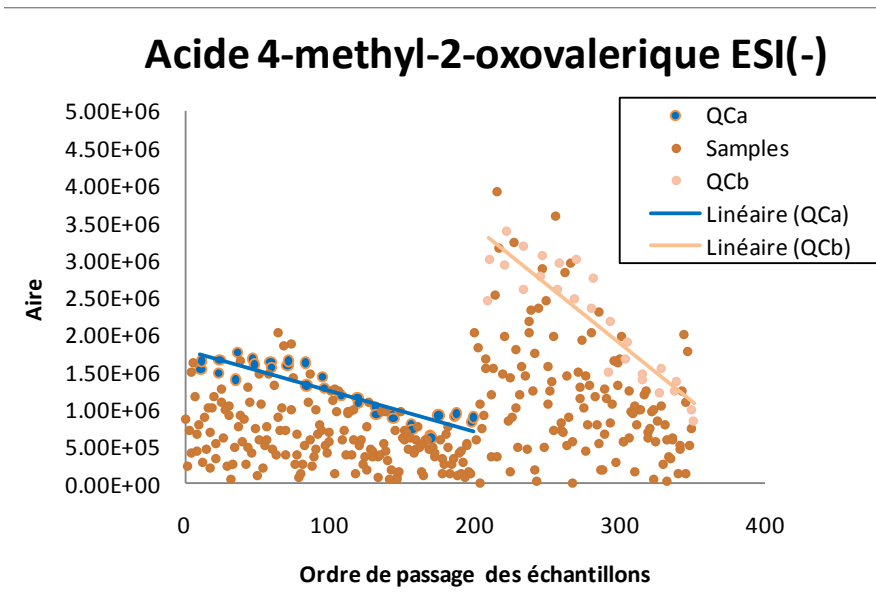
Ech bio
Ech bio
Ech bio

...

Ech bio
Ech bio

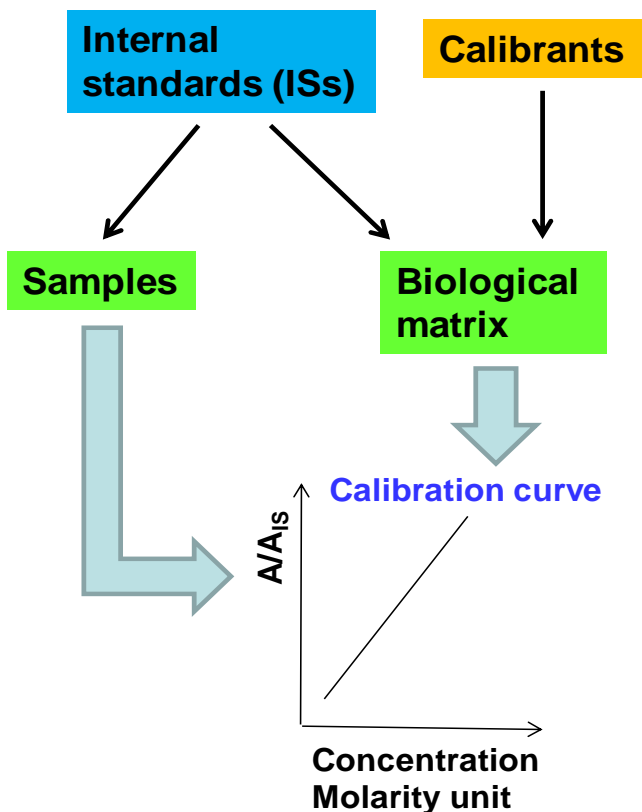
blanc
QC-pool
QC-pool + EI

10 échantillons bio



Quantitative metabolite profiling for large scale studies

Absolute quantification of metabolites



PLOS GENETICS

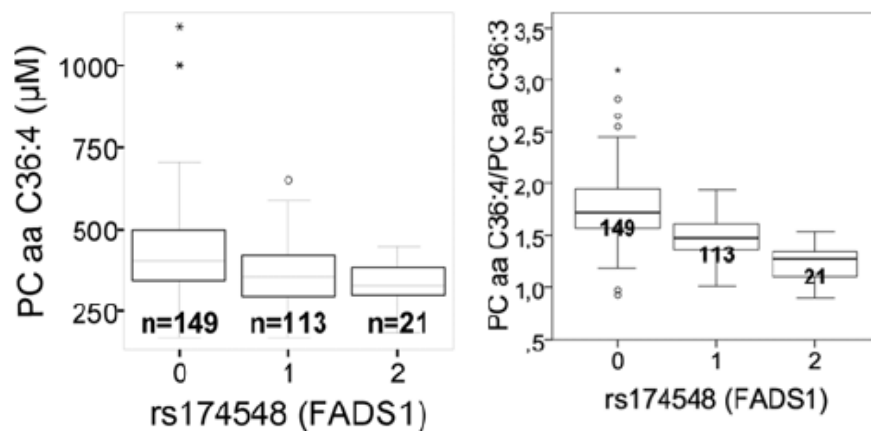
Genetics Meets Metabolomics: A Genome-Wide Association Study of Metabolite Profiles in Human Serum

Christian Gieger^{1,2}, Ludwig Geistlinger¹, Elisabeth Altmaier^{3,4}, Martin Hrabě de Angelis^{5,6}, Florian Kronenberg⁷, Thomas Meitinger^{8,9}, Hans-Werner Mewes^{3,10}, H.-Erich Wichmann^{1,2}, Klaus M. Weinberger¹¹, Jerzy Adamski^{5,6}, Thomas Illig¹, Karsten Suhre^{3,4*}

Quantitative measurement of 363 metabolites in 284 serum samples

«Genetically determined metabolotypes»

Polymorphism in the *FADS1* (fatty acid delta 5 desaturase) gene



Metabolomics (2012) 8:757–760
DOI 10.1007/s11306-012-0462-0

MetaboLights: towards a new COSMOS of metabolomics data management

Christoph Steinbeck · Pablo Conesa · Kenneth Haug · Tejasvi Mahendrakar ·
Mark Williams · Eamonn Maguire · Philippe Rocca-Serra · Susanna-Assunta Sansone ·
Reza M. Salek · Julian L. Griffin

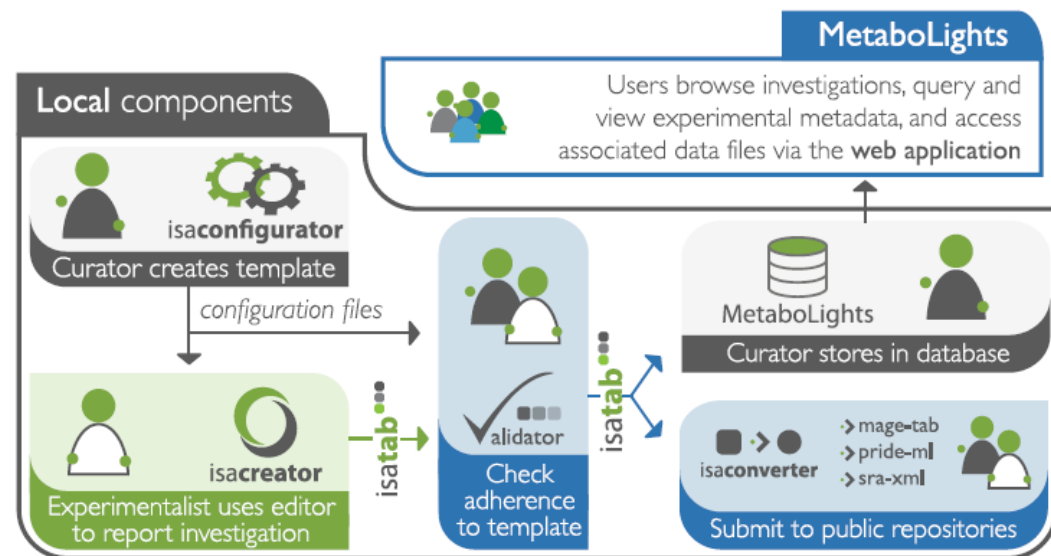
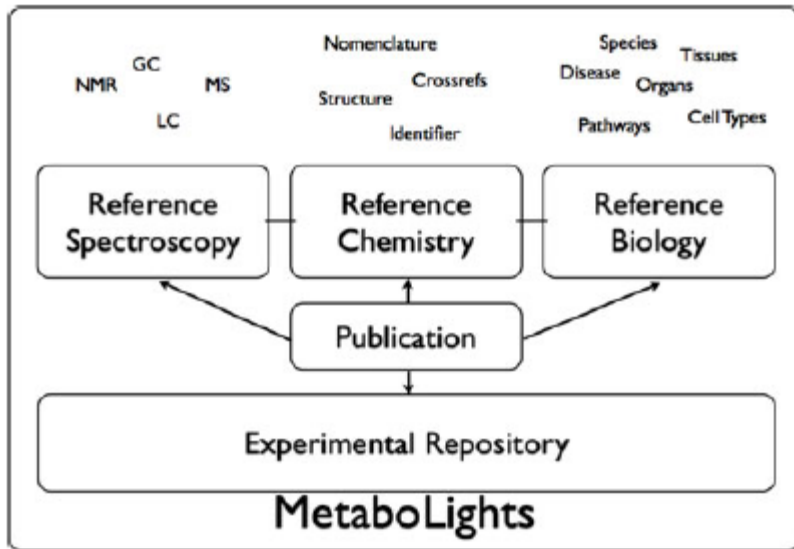


Fig. 2 MetaboLights data submission workflow

Conclusion

Apport significatif de la FT-MS dans la détection des métabolites (séparation des ions isobares) et dans leur identification (annotation/élucidation structurale).

Disponibilité de différents types de bases de données (bases spectrales de HRMS, de MS/MS, bases de données biochimiques et métabolomique). Les outils de normalisation et de quantification rendent possible la construction de bases de données de profils métaboliques.

Un des principaux enjeux: le partage des données concernant les composés inconnus. Pour cela, nécessité de standardisation des spectres MS/MS.

Très haute résolution (Orbitrap, FTICR) versus efficacité des acquisitions données dépendantes avec Q-TOF??

Remerciements

CEA/LEMM

Jean-François Heilier
Alexandra Lafaye
Céline Ducruix
Erwan Werner
Geoffrey Madalinski
Emmanuel Godat
Jérôme Cotton
Ying Xu
Aurélie Roux
Eric Ezan
Benoit Colsch
Samia Boudah

G.H. Pitié-Salpêtrière

Frédéric Sedel
Maria del Mar Amador
Fanny Mochel
Foudil Lamari

CEA/iRCM

Paul-Henri Roméo
Dhouha Darghouth
Lydie Oliveira
Bérengère Koehl
Marie-Françoise Olivier

Profilomic

Céline Ducruix, Jérôme Cotton, Simon
Broudin, Fanny Leroux, Bruno Corman,
Stéphanie Oursel, Victor Sabarly, Denis
Desoubzdanne

Laboratoire de Chimie Structurale Organique et Biologique (UPMC)

Jean-Claude Tabet,
Richard Cole
Denis Lesage, Sandra Alves
Estelle Paris, Farid Ichou

CEA/Programmes Transversaux de Technologies pour la santé et Tox Nuc

Jacques Grassi, Marie-Thérèse Ménager

CEA/DRT/LIST/LOAD

Antoine Souloumiac, Etienne
Thévenot,